FORM PTO-1390 TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DÓ/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371 INTERNATIONAL APPLICATION NO. INTERNATIONAL FILING DATE PCT/EP99/00478 January 27, January 30, 1998 TITLE OF INVENTION RECOMBINANT PROTEINS DERIVED FROM HGF AND MSP APPLICANT(S) FOR DO/EO/US MEDICO, Enzo; MICHIELI, Paolo; COLLESI, Chiara; CASELLI, Gianfranco; Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other JUL 2 6 2000 This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than de examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 3 A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date A copy of the International Application as filed (35 U.S.C. 371(c)(2)) is transmitted herewith (required only if not transmitted by the International Bureau). has been transmitted by the International Bureau. is not required, as the application was filed in the United States Receiving Office (RO/US). A translation of the International Application into English (35 U.S.C. 371(c)(3)). Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(2)). are transmitted herewith (required only if not transmitted by the International Bureau). b. A have been transmitted by the International Bureau. have not been made; however, the time limit for making such amendments has NOT expired. have not been made and will not be made. A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). Items 11. to 16. below concern document(s) or information included: An Information Disclosure Statement under 37 CFR 1.97 and 1.98.-1449 and International Search Report (PCT/ISA/210) w/ 6 references An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. A FIRST preliminary amendment. A SECOND or SUBSEQUENT preliminary amendment. A substitute specification. A change of power of attorney and/or address letter. 16. Other items or information: 1.) Nineteen (19) sheets of Formal Drawings 2.) Sequence Listing (8 pages) ~ 3.) International Preliminary Examination Report w/ 4 sheets amended claims ~ 4.) PCT Request (PCT/RO/101) ~

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/cqc July 26, 2000

528 Rec'd PCT/PTO 2 6 JUL 2000

IN THE U.S. PATENT AND TRADEMARK OFFICE

Applicant:

MEDICO, Enzo et al.

Int'l. Appl. No.:

PCT/EP99/00478

Appl. No.:

New

Group:

Filed:

July 26, 2000

Examiner:

For:

RECOMBINANT PROTEINS DERIVED FROM HGF AND MSP

PRELIMINARY AMENDMENT

#### BOX PATENT APPLICATION

Assistant Commissioner for Patents Washington, DC 20231

July 26, 2000

Sir:

The following Preliminary Amendments and Remarks respectfully submitted in connection with the above-identified application.

#### **AMENDMENTS**

### IN THE SPECIFICATION:

Please amend the specification as follows:

Before line 1, insert -- This application is the national phase under 35 U.S.C. § 371 of PCT International Application No. PCT/EP99/00478 which has an International filing date of January 27, 1999, which designated the United States of America. --

#### IN THE CLAIMS:

Please amend the claims as follows:

Claim 2: Line 1, change "claims 1-2" to --claim 1--

change "claims 1-2" to --claim 1 or 2--Claim 3: Line 1,

Claim 4: Line 1, change "claims 1-2" to --claim 1 or 2--

Claim 5: Line 2, change "1-5" to --1-2--

Claim 8: Line 1, change "claims 1-4" to --claim 1--

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Claim 10: Line 1, change "claims 1-4" to --claim 1--

Claim 11: Line 1, change "claims 1-4" to --claim 1--

Claim 13: Line 2, change "claims 1-4" to --claim 1--

#### REMARKS

The specification has been amended to provide a cross-reference to the previously filed International Application. The claims were also amended to delete improper multiple claims and to place the application into better form for examination. Entry of the present amendment and favorable action on the above-identified application are respectfully requested.

If necessary, the Commissioner is hereby authorized in this, concurrent, and future replies, to charge payment or credit any overpayment to Deposit Account No. 02-2448 for any additional fees required under 37 C.F.R. § 1.16 or under 37 C.F.R. § 1.17; particularly, extension of time fees.

Respectfully submitted,

BIRCH, STEWART, KOLASCH & BIRCH, LLP

Maymond C. Stewart, #21,066

P.O. Box 747

Falls Church, VA 22040-0747

(<del>7</del>03) 205-8000

(Rev. 04/19/2000)

Serial or Patent No.	MEDICO, Enzo et al.	
Filed or Issued:	09/600,991 July 26, 2000	Docket No.: 0471-0162P
or:	- RECOMBINANT PROTEINS DERIVE	D FROM HGF AND MSP
	VERIFIED STATEMENT (DECLARATION) CLAIMING (37 CFR 1.9(f) and 1.27(c)) - SMALL BUSIN	SMALL ENTITY STATUS
hereby declare that I ] the owner of the ] an official of the	am small business concern identified below: s small business concern empowered to act on b	pehalf of the concern identified below:
NAME OF CONCERN _ ADDRESS OF CONCER	DOMPE' S.p.A.  Via Campo di Pile L'AQUILA, Italy	
efined in 13 CFR 121.3 ection 41(a) and (b) c ncluding those of its cumber of employees of the persons employed on iscal year, and (2) con ontrols or has the powentrol both.	ne above-identified small business concern questions. As, and reproduced in 37 CFR 1.9(d), for possible of Title 35, United States Code, in that the affiliates, does not exceed 500 persons. For the business concern is the average over the last a full-time, part-time or temporary basis and a full-time, part-time or temporary basis are affiliates of each other when either to control the other, or a third party or ghts under contract or law have been conveyed.	purposes of paying reduced fees under a number of employees of the concern, or purposes of this statement, (1) the previous fiscal year of the concern of during each of the pay periods of the cr, directly or indirectly, one concern parties controls or has the power to
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escribed in [ ] [ ] [ ]		, filed
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# RECOMBINANT PROTEINS DERIVED FROM HGF AND MSP

### Field of the invention

The present invention relates to recombinant proteins obtained from the combination of structural domains derived from the  $\alpha$  subunits of hepatocyte growth factor (HGF) and macrophage stimulating protein (MSP).

In particular, the engineered factors of the invention are obtained by combination of the hairpin loop and kringle domains of the α chains of HGF and/or MSP, so as to obtain a structure having two superdomains with an intervening linker sequence. Moreover, the invention relâtes to DNA sequences encoding the above mentioned recombinant proteins, to the expression vectors comprising said DNA sequences and to host cells containing said expression vectors. The recombinant proteins of the present invention are biologically active, and their activity can be measured by determination of their ability to induce activation of the Met tyrosine kinase receptor, their "scattering" effect on epithelial cells, and their protective effect against cell death induced by chemotherapic drugs (vide infra). Therefore, these molecules can conveniently be used to prevent or treat the toxic side effects of the chemotherapeutical treatment of tumours, and to reduce introgenic cell damage induced by other types of drugs.

## Technological background

Hepatocyte Growth Factor (HGF) and Macrophage Stimulating Protein (MSP) are highly related proteins both structurally and



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functionally (Fig. 1 and 2). Both these factors are secreted as an inactive precursor, which is processed by specific proteases which recognise a cleavage site inside the molecule, dividing the protein in two subunits. These subunits, named α chain and β chain, are linked by a disulphide bond. Thus, the mature factor is an α-β dimeric protein. Only the mature (dimeric) form of the factor is able to activate its receptor at the surface of the target cells (the Met tyrosine kinase in the case of HGF and the Ron tyrosine kinase in the case of MSP) and therefore to mediate biological responses (Naldini, L. et al., 1992, EMBO J. 11: 4825-4833; Wang, M. et al., 1994, J. Biol. Chem. 269; 3436-3440; Bottaro, D. et al., 1991, Science 25: 802-804; Naldini, L. et al., 1991, EMBO J. 10: 2867-2878; Wang, M. et al., 1994, Science 266: 117-119; Gaudino, G. et al., 1994, EMBO J. 13: 3524-3532).

The  $\alpha$  chain of both factors contains a hairpin loop (HL) structure and four domains with a tangle-like structure named kringles (K1-K4; Nakamura T et al., 1989, Nature 342:440-443; Han, S. et al.., 1991, Biochemistry 30: 9768-9780). The precursor also contains a signal sequence (LS) of 31 amino acids (in the case of HGF) or of 18 amino acids (in the case of MSP), removed in rough endoplasmic reticulum, which directs the neoformed peptide to the secretive pathway. The  $\beta$  chain contains a box with a sequence homologous to that typical of serine proteases, but it has no catalytic activity (Nakamura T et al., 1989, Nature 342:440-443; Han, S. et al.., 1991, Biochemistry 30: 9768-9780). Both  $\alpha$  and  $\beta$  chains contribute to the binding of the growth factor to the respective receptor (Met for HGF and

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**(1)** 

Ron for MSP).

HGF and MSP polypeptides are able to induce a variety of biological effects besides cell proliferation. The main biological activities of these molecules are: stimulation of cell division (mitogenesis); stimulation of motility (scattering); induction of polarisation and cell differentiation; induction of tubule formation (branched morphogenesis); increase of cell survival (protection from apoptosis). The tissues that respond to HGF and MSP stimulation are those where cells express the respective Met (HGF) and Ron (MSP) receptors. The most important target tissues of these factors are epithelial cells of different organs, such as liver, kidney, lung, breast, pancreas and stomach, and some cells of the hematopoietic and nervous systems. A detailed review of the biological effects of HGF and MSP in the various tissues can be found in Tamagnone, L. & Comoglio, P., 1997, Cytokine & Growth Factor Re-views, 8: 129-142, Elsevier Science Ltd.; Zarnegar, R. Michalopoulos, G., 1995, J. Cell Biol. 129: 1177-1180; Medico, E. et al., 1996, Mol. Biol. Cell, 7: 495-504; Banu, N. et al., 1996, J. Immunol. 156: S2933-2940.

In the case of HGF, the hairpin loop and the first two kringles are known to contain the sites of direct interaction with the Met receptor (Lokker NA et al., 1992, EMBO J., 11:2503-2510; Lokker, N. et al., 1994, Protein Engineering 7: 895-903). Two naturally-occurring truncated forms of HGF produced by some cells by alternative splicing have been described. The first one comprises the first kringle (NK1-HGF Cioce, V. et al., 1996, J. Biol. Chem.

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271: 13110-13115) whereas the second one spans to the second kringle (NK2-HGF Miyazawa, K. et al., 1991, Eur. J. Biochem. 197: 15-22). NK2-HGF induces cell scattering, but it is not mitogenic as the complete growth factor is (Hartmann, G. et al., 1992, Proc. Natl. Acad. Sci. USA 89: 11574-11578). However, NK2-HGF regains, mitogenic activity in the presence, of heparin, glucosaminoglycan that binds HGF through a domain contained in the first kringle and which is likely to induce dimerization of NK2-HGF (Schwall, R. et al., 1996, J. Cell Biol. 133: 709-718). Moreover NK2-HGF, being a partial agonist of Met, behaves as a competitive inhibitor of HGF as far as the mitogenic activity is concerned (Chan, A. et al., 1991, Science 254: 1382-1385). NK1-HGF has also been described to exert partial stimulation of Met and competitive inhibition of HGF mitogenic activity (Cioce, V. et al., 1996, J. Biol. Chem. 271: 13110-13115). Anyway, a truncated factor is endowed with an activity markedly lower than the recombinant factors described in the invention, as shown in example 3.

In the case of MSP, the interaction sites with the Ron receptor are less understood: some preliminary studies suggest a situation opposite of that of HGF, i.e. the  $\beta$  chain directly binds the receptor whereas the  $\alpha$  chain would act stabilizing the complex (Wang MH et al., 1997, J. Biol. Chem. 272:16999-17004).

The therapeutical use of molecules such as HGF and MSP is potentially valuable in a wide range of pathologies (Abdulla, S., 1997, Mol. Med. Today 3: 233). Nevertheless, a number of technical

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as well as biological complications make the application of these molecules in clinics difficult. First of all, the pleiotropic character of these factors can causes poorly selective biological responses, which involve undesired side effects. For example, the use of HGF to prevent some side effects of the chemotherapeutic drug cisplatin has been proposed (Kawaida K et al., 1994, Proc. Natl. Acad. Sci. 91:4357-4361). Cancer patients treated with this drug can suffer kidney acute damage due to the cytotoxic action of cisplatin on proximal tubule epithelial cells. HGF is able to protect these cells against programmed death (apoptosis) induced by cisplatin, but at the same time it can induce an undesired proliferation of neoplastic cells. Other problems related to the pharmaceutical use of HGF and MSP are the necessity of their proteolytic activation and their stability, which causes technical problems. The NK1 and NK2 truncated forms of HGF do not require proteolytic activation, but they have a reduced biological activity.

### Summary of the invention

recombinant molecules provides The invention present composed of a combination of structural domains derived from the α chains of HGF and/or MSP, which overcome the problems of the prior art molecules described above. The molecules of this invention are composed of two superdomains connected by a linker. Each superdomain is composed of a combination of the HL and K1-K4 domains of the a chain of HGF and/or MSP. These engineered not require factors induce selective biological responses, do

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proteolytic activation, are stable and are more active than the truncated forms of HGF described previously.

#### Detailed disclosure of the invention

The present invention relates to recombinant proteins (which will be hereinafter referred to indifferently as proteins, molecules, engineered or recombinant factors) characterised by a structure that comprises two superdomains, each consisting of a combination of HL and K1-K4 domains derived from the α chain of HGF and/or MSP, linked by a spacer sequence or a linker. In particular, the invention relates to proteins of general formula (I)

$$[A] - B - [C] - (D)_y$$
 (I)

in which

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[A] corresponds to the sequence (LS)<sub>m</sub>-HL-K1-(K2)<sub>n</sub>-(K3)<sub>o</sub>-(K4)<sub>p</sub> wherein (the numbering of the following amino acids refers to the HGF and MSP sequences as reported in Fig. 1 and 2, respectively):

LS is an amino acid sequence corresponding to residues 1-31 of HGF or 1-18 of MSP;

HL is an amino acid sequence derived from the  $\alpha$  chain of HGF starting between residues 32-70 and ending between residues 96-127; or it is an amino acid sequence derived from the  $\alpha$  chain of MSP starting between residues 19-56 and ending between residues 78-109;

K1 is an amino acid sequence derived from the  $\alpha$  chain of HGF starting between residues 97-128 and ending between residues 201-205; or it is an amino acid sequence derived from the  $\alpha$  chain of MSP starting between residues 79-110 and ending between residues 186-190;

K2 is an amino acid sequence derived from the  $\alpha$  chain of HGF starting between residues 202-206 and ending between residues 283-299; or it is an amino acid sequence derived from the  $\alpha$  chain of MSP starting between residues 187-191 and ending between residues 268-282;

5 K3 is an amino acid sequence derived from the α chain of HGF starting between residues 284-300 and ending between residues 378-385; or it is an amino acid sequence derived from the α chain of MSP starting between residues 269-283 and ending between residues 361-369;

K4 is an amino acid sequence derived from the α chain of HGF starting between residues 379-386 and ending between residues 464-487; or it is an amino acid sequence derived from the α chain of MSP starting between residues 362-370 and ending between residues 448-481;

m, n, o, p can be 0 or 1;

 $n \ge o \ge p$ ;

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the sum n + o + p is an integer from 1 to 3 or 0, with the proviso that

B is the sequence  $[(X)_q Y]_r$ , wherein X = Gly and Y = Ser, or Cys, or Met, or Ala;

q is an integer from 2 to 8;

r is an integer from 1 to 9;

[C] corresponds to the sequence HL-K1-(K2)<sub>s</sub>-(K3)<sub>t</sub>-(K4)<sub>u</sub> wherein HL, K1-K4 are as defined above,

s, t, u are 0 or 1; the sum s + t + u is an integer from 1 to 3 or 0, with the proviso that  $s \ge t \ge u$ ;

D is the sequence W-Z, wherein W is a conventional proteolytic site, Z is any tag sequence useful for the purification and detection of the protein; y

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is 0 or 1.

Non-limiting examples of W are consensus sequences for enterokinase protease, thrombin, factor Xa and IgA protease.

Preferred proteins of general formula (I), are those in which: the HL domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 32 to 127, or a sequence of MPS  $\alpha$  chain ranging from amino acids 19 to 98; the K1 domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 128 to 203, or a sequence of MPS  $\alpha$  chain ranging from amino acids 99 to 188; the K2 domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 204 to 294, or a sequence of MPS  $\alpha$  chain ranging from amino acids 189 to 274; the K3 domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 286 to 383, or a sequence of MPS  $\alpha$  chain ranging from amino acids 275 and 367; the K4 domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 384 to 487, or a sequence of MPS  $\alpha$  chain ranging from amino acids 384 to 487, or a sequence of MPS  $\alpha$  chain ranging from amino acids 368 and 477.

Among the possible combinations of the domains of general formula (I), the following (II) and (III) are preferred, concerning two recombinant factors named Metron Factor-1 and Magic Factor-1, respectively:

LS<sub>MSP</sub>-HL<sub>MSP</sub>-K1<sub>MSP</sub>-K2<sub>MSP</sub>-L-HL<sub>HGF</sub>-K1<sub>HGF</sub>-K2<sub>HGF</sub>-D (Metron Factor-1)
(II)

and

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 $LS_{HGF}-HL_{HGF}-K1_{HGF}-K2_{HGF}-L-HL_{HGF}-K1_{HGF}-K2_{HGF}-D \quad (Magic \quad Factor-1)$  (III)

For both molecules, L is a linker sequence (Gly<sub>4</sub>Ser)<sub>3</sub>, D is a tag

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sequence Asp<sub>4</sub>-Lys-His<sub>6</sub>.

For Metron Factor-1, LS<sub>MSP</sub> is the sequence 1-18 of MSP,  $HL_{MSP}$  is the sequence 19-56 of MSP,  $K1_{MSP}$  is the sequence 99-188 of MSP,  $K2_{MSP}$  is the sequence 189-274 of MSP,  $HL_{HGF}$  is the sequence 32-127 of HGF,  $K1_{HGF}$  is the sequence 128-203 of HGF,  $K2_{HGF}$  is the sequence 204-294 of HGF.

For Magic Factor-1,  $HL_{HGF}$ ,  $K1_{HGF}$ ,  $K2_{HGF}$  are as defined above,  $LS_{HGF}$  is the sequence 1-31 of HGF.

The hybrid molecules of the invention are prepared by genetic engineering techniques according to a strategy involving the following steps:

- a) construction of DNA encoding the desired protein;
- b) insertion of DNA in an expression vector;
- c) transformation of a host cell with recombinant DNA (rDNA);
- culture of the transformed host cell so as to express the recombinant protein;
  - e) extraction and purification of the produced recombinant protein.

The DNA sequences corresponding to HGF or MSP structural domains can be obtained by synthesis or starting from DNA encoding for the two natural factors. For example, screening of cDNA libraries can be carried out using suitable probes, so as to isolate HGF or MSP cDNA. Alternatively, HGF or MSP cDNA can be obtained by reverse transcription from purified mRNA from suitable cells.

cDNAs coding for the fragments of HGF and MSP β chains can be amplificated by PCR (Mullis, K.B. and Faloona, F.A., 1987, Methods in

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Enzymol. 155, 335-350), and the amplification products can be recombined making use of suitable restriction sites, naturally occurring in the factor sequences or artificially introduced in the oligonucleotide sequence used for the amplification.

In greater detail, one of the above mentioned strategies can be the following:

the portions of DNA encoding the LS, HL, K1, K2, K3 and K4 domains are amplificated by PCR from HGF or MSP cDNA and then recombined to corresponding to [A] and [C]. sequences hvbrid the obtain Oligonucleotides recognising sequences located at the two ends of the domains to be amplificated are used as primers. Primers are designed so as to contain a sequence allowing recombination between the DNA of a domain and the adjacent one. Said recombination can be carried out by endonuclease cleavage and subsequent ligase reaction, or making use of the recombinant PCR method (Innis, NA et al., 1990, in PCR Protocols, Academic Press, 177-183).

The sequence encoding the domain B (linker) can be obtained by synthesis of a double chain oligonucleotide, which can be inserted between [A] and [C] using suitable restriction sites.

The resulting three fragments encoding for [A], [B] and [C] are then inserted in the correct sequence in a suitable vector. In this step it can be decided whether to add or not the domain D (tag), obtained by synthesis analogously to domain B, downstream fragment [C].

The recombinant expression vector can contain, in addition to the recombinant construct, a promoter, a ribosome binding site, an initiation

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codon, a stop codon, optionally a consensus site for expression enhancers.

The vector can also comprise a selection marker for isolating the host cells containing the DNA construct. Yeast or bacteria plasmids, such as plasmids suitable for Escherichia Coli, can be used as vectors, as well as bacteriophages, viruses, retroviruses, or DNA.

The vectors are cloned preferably in bacterial cells, for example in Escherichia Coli, as described in Sambrook J., 1989, Molecular Cloning, Cold Spring Harbor Laboratory Press, New York, and the colonies can be selected, for example, by hybridisation with radiolabelled oligonucleotide probes; subsequently, the rDNA sequence extracted from the positive colonies is determined by known methods.

The vector with the recombinant construct can be introduced in the host cell according to the competent cell method, the protoplast method, the calcium phosphate method, the DEAE-dextran method, the electric impulses method, the in vitro packaging method, the viral vector method, the micro-injection method, or other suitable techniques.

Host cells can be prokaryotic or eukaryotic, such as bacteria, yeasts or mammal cells, and they will be such as to effectively produce the recombinant protein.

After transformation, cells are grown in a suitable medium, which can be for example MEM, DMEM or RPMI 1640 in the case of mammal host cells.

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The recombinant protein is secreted in the culture medium from which it can be recovered and purified with different methods, such as mass exclusion, absorption, affinity chromatography, salting-out, precipitation, dialysis, ultrafiltration.

A simple, rapid system for the production of the molecules of the invention is, for example, transient expression in mammal cells.

Accordingly, the plasmid containing the recombinant DNA fragment, for example PMT2 (Sambrook, J. et al., 1989, Molecular Cloning, Cold Spring Harbor Laboratory Press), is transfected in suitable recipient cells, such as Cos7 (Sambrook, J. et al., supra) by the calcium phosphate technique or other equivalent techniques. Some days after transfection, the conditioned medium of the transfected cells is collected, cleared by centrifugation and analysed for its content in factor. For this analysis, antibodies directed against HGF or MSP, or against any tag sequence, can be used: the supernatant is immunoprecipitated and then analysed by western blot with the same antibody. The supernatant containing the recombinant factor can also be used directly for biochemical and biological tests. The protein can be purified, for example, using a poly-histidine tag sequence, by absorption on a nickel resin column and subsequent elution with imidazole.

The biochemical properties of the recombinant factors of the invention were tested in connection with their ability to activate Met and Ron receptors.

Sub-micromolar concentrations of the factors have proved to induce phosphorylation in Met tyrosine in human epithelial cells A549, whereas

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they do not induce phosphorylation above basal values in cells expressing Ron. On the whole, the tests proved that the first two kringles of HGF maintain their ability to interact and to activate Met tyrosine kinase receptor, whereas the corresponding first two kringles of MSP are not sufficient for modulating the catalytic activity of the Ron receptor. However, the interaction with Ron, although at low affinity, can contribute to the recruitment of the factor at the cell surface, playing a similar role to low affinity receptors (of mature glycoprotein) which recruit the HGF intact molecule through the heparin-binding domain.

The molecules of the invention have a marked biological activity, measured by the scattering tests, and a protecting activity against cell apoptosis induced by cisplatin or etoposide.

In particular, the supernatant containing the recombinant factor has been found to promote scattering of epithelial cells of various nature even at nanomolar concentrations. In these tests, kidney epithelial cells (MDCK) or hepatocyte precursors (MLP29) were used.

In an in vitro experimental system, in which DNA fragmentation typical of apoptotic cells is evaluated by the TUNEL method (Gavrieli, Y. et al., 1992, J. Cell. Biol. 117, 493-501), the recombinant factors protect against apoptosis induced by chemotherapeutic drugs at levels comparable with HGF and remarkably higher than MSP. The engineered molecules proved to be active on human primary epithelial cells from proximal tubule (PTECs), on an immortalised PTECs line (Loc) and on the already cited murine hepatocytes MLP29.

Among the applications of the recombinant molecules of the

invention, the following can be cited:

- prevention of myelotoxicity; in particular they can be used for the expansion of marrow precursors, to increase proliferation of the hematopoietic precursors or to stimulate their entry in circle;
- prevention of liver and kidney toxicity, and of mucositis following antineoplastic treatments; in particular the recombinant factors can be used to prevent toxicity (apoptosis) on differentiated cell elements of liver, kidney and mucosa of the gastroenteral tract, and to stimulate staminal elements of cutis and mucosas to allow the regeneration of germinative layers;
  - prevention of chemotherapeutic neurotoxicity.

In general, the proteins of the invention provide the following advantages, compared with the parent molecules HGF and MSP:

- they are smaller molecules with a more compact structure;
- they are more stable and are produced in higher amounts;
  - they require no endoproteolytic cleavage for activation, which transforms the HGF and MSP precursors into the respective active forms;
- they can be engineered in combinations of different functional domains, thereby modulating the biological effects, increasing the favourable ones and reducing those undesired (for example, protection from apoptosis versus cell proliferation).

The invention has to be considered also directed at amino acid and nucleotide sequences referred to formula (I), having modifications which can, for example, derive from degeneration of genetic code, without

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therefore modifying the amino acid sequence, or from the deletion, substitution, insertion, inversion or addition of nucleotides and/or bases according to all the possible methods known in the art.

Furthermore, the invention relates to the expression vectors comprising a sequence encoding for a protein of general formula (I), which can be plasmids, bacteriophages, viruses, retroviruses, or others, and to host cells containing said expression vectors.

Finally, the invention relates to the use of the recombinant proteins as therapeutical agents, and to pharmaceutical compositions containing an effective amount of the recombinant proteins together with pharmacologically acceptable excipients.

# Description of the Figures

(In the following legends, -His located after the name of the parent factors, truncated or recombinant, or of the plasmids, means that the respective sequences contain a poly-histidine tag).

### Figure 1:

- a) Nucleotide and amino acid sequence of human HGF (Gene Bank # M73239; Weidner, K.M., et al., 1991, Proc. Acad. Sci. USA, 88:7001-7005). In contrast to the cited reference, in the numbering used herein, nucleotide No. 1 is the first base of the initiation codon (the A of the first ATG). The first amino acid is the corresponding methionine. The cDNA untranslated regions at 5' and 3' are neither represented nor considered in the numbering.
- b) Nucleotide and amino acid sequence of human MSP (Gene Bank # L11924; Yoshimura, T., et al., 1993, J. Biol. Chem., 268:15461-

15468). In contrast to the cited reference, in the numbering used herein nucleotide No. 1 is the first base of the initiation codon (the A of the first ATG). The first amino acid is the corresponding methionine. The cDNA untranslated regions at 5' and 3' are neither represented nor considered in the numbering.

## Figure 2:

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- a) Molecular structure of Metron Factor-1. The leader sequence is removed from the cells used for the production before secretion and is therefore absent in the mature molecule. The poly-histidine tag can be removed by digestion with the protease enterokinase.
- b) Nucleotide and amino acid sequence of Metron-Factor-1. The nucleotide sequence starts with the EcoRI site and terminates with the Sall site (first six bases and last six bases, respectively). The initiation codon (ATG) and the stop codon (TAG) are underlined.

## Figure 3:

- a) Molecular structure of Magic Factor-1. The leader sequence is removed from the cells used for the production before secretion and is therefore absent in the mature molecule. Poly-histidine tag can be removed by digestion with the protease enterokinase.
- b) Nucleotide and amino acid sequence of Magic Factor-1. The nucleotide sequence starts with the SalI site (first six bases and last six bases, respectively). The initiation codon (ATG) and the stop codon (TAG) are underlined.

### Figure 4:

Production of Metron-F-1 by transient transfection of mammal

cells. The conditioned supernatants from BOSC cells transfected with the control plasmid (CTRL) or with pRK7-Metron F-1-His were immunoprecipitated with an anti-MSP polyclonal antibody and detected by western blot with the same antibody.

# 5 Figure 5:

Quantitation of the recombinant proteins by western blot. (A) The proteins were absorbed on Sepharose-A-heparin beads and detected with an anti-poly-histidine monoclonal antibody. (B) The proteins were immunoprecipitated with an anti-MSP polyclonal antibody and detected with an anti-poly-histidine monoclonal antibody.

## Figure 6:

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Scattering test carried out on kidney epithelial cells (MDCK) using the recombinant proteins prepared by transient transfection. The protein content was quantified by western blot (see Fig. 5). (A) non-stimulated cells; (B) cells stimulated with control supernatant; (C) cells stimulated with HGF-His; (D) cells stimulated with NK2-HGF-His; (E) cells stimulated with Metron Factor-1; (F) cells stimulated with Magic Factor-1.

# Figure 7:

Activation (phosphorylation) of Met receptor by the hybrid factor Metron Factor-1. Human epithelial cells (A549) were stimulated with supernatants conditioned from BOSC cells transfected with the control plasmid (CTRL) or with pRK7-Metron-F-1-His (METRON F-1) at the indicated dilutions. Cell lysates from the stimulated cells were immunoprecipitated with an anti-Met monoclonal antibody and detected by western blot with an anti-phosphotyrosine monoclonal antibody.

## Figure 8:

Protective effect of Metron-F-1 against acute renal failure induced by HgCl<sub>2</sub> in vivo. Balb-c mice were injected i.v. with Metron-F-1 or vehicle at 0.5 h before and 6, 12, 24, 36 and 48 h after HgCl<sub>2</sub> i.v. administration. BUN and histological evaluation of renal necrosis were measured at 72 h.

Data expressed as mean + e.s. of 7 animals/group (BUN) or 3 animals/group (histology).

The following examples illustrate in greater detail the invention.

# 10 Example 1a: Preparation of the recombinant construct encoding Metron Factor-1

HGF cDNA was obtained by the RT-PCR technique (Reverse Transcriptase PCR; in: Innis, M. A., et al., 1990, PCR Protocols, Academic Press, 21-27) from a human lung fibroblast cell line (MRC5; Naldini, L. et al., 1991, EMBO J. 10: 2867-2878). MSP cDNA was obtained with the same technique from human liver (Gaudino, G., et al., 1994, EMBO J. 13: 3524-3532).

The fragment corresponding to MSP LS-HL-K1-K2 was amplified by PCR using MSP cDNA as template and the following oligonucleotides as primers:

P1 (sense)

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- 5' CGCGCGGAATTCCACCATGGGGTGGCTCCCACTCCT 3'
- P2 (antisense)
- 5' CGCGCGCTCGAGGCGGGGCTGTGCCTCGGACCCGCA 3'
- in which the underlined palindromic sequences are the restriction sites for

the enzymes EcoRI (oligonucleotide P1) and XhoI (oligonucleotide P2). The PCR product was digested with the restriction enzymes EcoRI and XhoI and then purified by electrophoresis on agarose gel.

The fragment corresponding to HL-K1-K2 of HGF was amplified by PCR using HGF cDNA as template and the following oligonucleotides as primers:

P3 (sense)

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5' CGCGCG<u>TCTAGA</u>GGGACAAAGGAAAAGAAGAAATAC 3' P4 (antisense)

5' CGCGCGAAGCTTTGTCAGCGCATGTTTTAATTGCAC 3' in which the underlined palindromic sequences are the restriction sites for the enzymes XbaI (oligonucleotide P3) and HindIII (oligonucleotide P4). The PCR product was digested with the restriction enzymes XbaI and HindIII and then purified by electrophoresis on agarose gel.

For the linker sequence, the following partially complementary oligonucleotides were synthesised, and were subsequently annealed to obtain a double strand DNA fragment with sticky ends:

P5 (sense)

5'<u>TCGA</u>GGCGGTGGCGGTTCTGGTGGCGGTGGCTCCGGCGGTGGCGGTTCT3'

20 P6 (antisense)

5'CTAGAGAACCGCCACCGCCGGAGCCACCGCCACCAGAACCGCCACCGCCC3' in which the underlined bases are the sequences compatible with the restriction sites for the enzymes XhoI (oligonucleotide P5) and XbaI (oligonucleotide P6).

The resulting three DNA fragments were subcloned in the EcoRI-

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HindIII sites of the expression vector pRK7 (Gaudino, G., et al., 1994, EMBO J. 13: 3524-3532), to obtain the recombinant plasmid pRK7-Metron-F-1, containing all the components of Metron Factor-1 except the tag sequence.

For the insertion of the tag sequence, the following partially complementary oligonucleotides were synthesised, and were subsequently annealed to obtain a double strand DNA fragment with sticky ends:

P7 (sense)

5' AGCTGACGACGACAAACACCACCACCACCACCACCACTAGGGTCGAC'3'

10 P8 (antisense)

5'AGCTGTCGACCCTAGTGGTGGTGGTGGTGGTGGTGTTTGTCGTCGTCGTC3' in which the underlined bases are compatible with the HindIII restriction site and the boxed palindromic sequences are the consensus sequences for the enzyme Sall. The resulting double strand DNA fragment was inserted in the restriction site HindIII of the recombinant plasmid obtained at the previous step (destroying the HindIII site and creating the Sall site), to obtain the plasmid pRK7-Metron-F-1-His.

# **Example 1b:** Production of Metron Factor-1

The expression vector pRK7 contains a promoter of human cytomegalovirus immediate-early gene (CMV) and an episomal replication origin site of the DNA virus SV40. Therefore, this plasmid is particularly suitable for the expression of proteins in cells expressing the large T antigen of the virus SV40, such as kidney epithelial BOSC cells (Sambrook, J. et al.., 1989, Molecular Cloning, Cold Spring Harbor Laboratory Press). Metron Factor-1 can then be produced by transient

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transfection of plasmid pRK7-Metron F-1-His in BOSC cells.

For transfection, 10<sup>6</sup> cells are seeded at day 0 in a 100 mm plate in 90% Dulbecco's Modified Eagle Medium (DMEM)-10% bovine calf serum (10 ml/plate). At day 1, cells are transfected with 10 µg/plate of pRK7-Metron-F-1-His by lipofection, using the protocol provided by the lipofectin producer (Gibco-BRL). At day 2, the DNA-containing medium is substituted by fresh medium with low content in serum (99.5% DMEM-0.5% bovine calf serum). At day 4 (48 hours after the end of the transfection), the medium is collected, cleared by centrifugation, and analysed for its content in Metron Factor-1.

This analysis can be carried out in different ways. For example, the recombinant protein present in the cleared supernatant can be immunoprecipitated with an anti-MSP antibody and then detected by western blot with the same antibody (Fig. 4). In the example shown in figure 4, 500 µl of supernatant (cleared by centrifugation, buffered in 25 mM HEPES and added with a protease inhibitors cocktail) were immunoprecipitated (2 hours at 4° C) with 20 µl of Sepharose-A beads (Pharmacia) covalently conjugated with 2 µl of anti-MSP polyclonal antibody. The beads pellet was washed 3 times with 500 µl of washing buffer (20 mM HEPES pH 7.4; 150 mM NaCl; 0.1% Triton X-100; 10% glycerol) and heated at 90° C for 2 minutes in 100 µl of Laemmli buffer. Eluted proteins were separated by SDS-PAGE on 8% BIS-acrylamide gel, transferred onto membrane (Hybond-C; Amersham) and analysed by western blot. For this analysis, the same rabbit serum used for immunoprecipitation was employed as primary antibody with a 1:1000

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dilution and protein A conjugated with peroxidase (Amersham) was used as secondary antibody. Protein A was detected by ECL (Amersham) following the protocol provided by the producer.

Alternatively, the recombinant protein can be partially purified by adsorption on Sepharose-A beads conjugated with heparin and subsequent analysis by western blot using antibodies directed to poly-histidine tag (Fig. 5).

In the example shown in figure 5, the Sepharose-A-heparin beads (20 µl; Pierce) were incubated (4 hours at 4° C) with 500 µl of supernatant (cleared by centrifugation, buffered in 25 mM HEPES and added with a protease inhibitors cocktail) in the presence of 500 mM NaCl, washed with suitable buffer (500 mM NaCl; 20 mM HEPES pH 7.4; 0.1% Triton X-100; 10% glycerol) and heated at 90° C for 2 minutes in 100 µl Laemmli buffer. Eluted proteins were separated by SDS-PAGE on 8% bisacrylamide gel, transferred onto membrane (Hybond-C; Amersham) and analysed by western blot. For this analysis, a mouse monoclonal antibody to poly-histidine (Invitrogen) diluted 1:5000 was used as primary antibody and an anti-mouse IgG ovine antibody conjugated with peroxidase (Amersham) was used as secondary antibody. The secondary antibody was detected by ECL (Amersham) following the protocol provided by the producer.

The procedure of adsorption on heparin beads can also be used as protocol for the semi-purification of the recombinant protein. Furthermore, the molecule can additionally be purified making use of the poly-histidine affinity to heavy metals such as nickel. The protein containing poly-

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histidine tag can be adsorbed on a nickel resin column (Invitrogen) and subsequently eluted with imidazole (the detailed protocol is provided by the manufacturer).

## Example 1c: METRON-F-1 production in insect cells

The cDNA encoding for Metron-F1 was subcloned in a suitable expression vector (p-FASTBAC) to generate a recombinant plasmid containing the Metron-F1 gene (p-FASTBAC-Metron). A competent E. Coli strain (DH10 Bac) was transformed with p-FASTBAC-Metron to generate BACMID DNA. The DNA of positive colonies was isolated and checked by PCR to show the correct integration of the expression vector. Subsequently, the DNA from three clones was transfected into Sf9 insect cells with CellFECTIN reagent to produce virus particles. Virus titer was tested by a plaque assay. Single plaques were isolated and used for further propagation of the baculovirus. Viral stock was subsequently expanded in insect cells to scale up METRON-F-1 production. To verify protein expression, insect cells were infected with a multiplicity of infection (MOI) of 1 in a small-scale reactor. Samples of supernatants were analysed by SDS-PAGE followed by western blotting.

To produce amounts adequate for in vivo testing, insect cells were propagated in a 2.5-Liter stirred tank bioreactor. Cells were grown to a cell density of 1.106 ml<sup>-1</sup> before they were infected with a MOI of 1. Cell suspension was harvested 3 days post infection. The supernatant containing the recombinant protein was separated by centrifugation. The presence of Metron F-1 in the supernatant was proved by SDS-PAGE followed by western blotting. Metron F-1 was pre-purified by a dual step

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affinity chromatography on heparin sepharose (heparin-Hi Trap, Pharmacia) at 6° C. For in vivo testing or for further purification steps, the eluted fractions containing Metron F-1 were desalted by Sephadex G-25 chromatography (PD-10 or HiPrep 26/10, Pharmacia). Metron F-1 was further purified by chromatography on HisTrap columns (Pharmacia) and eluted by an imidazole gradient (0-0.5 M) using either a low-pressure system (Econo System, BIO-RAD) or an FPLC system (Pharmacia). Metron F-1 was eluted at an imidazole concentration of about 0.15 M. For in vivo testing, the eluted fractions containing Metron F-1 were freed of imidazole by Sephadex G-25 chromatography as already described, using the buffer to be used for animal treatment.

# Example 2a: Preparation of the recombinant construct encoding for Magic Factor-1

HGF cDNA and the plasmid pRK7-Metron-F-1-His described above were used as starting DNA. The fragment corresponding to LS-HL-K1-K2 of HGF was amplificated by PCR using HGF cDNA as template and the following oligonucleotides as primers:

P9 (sense)

5' CGCGCGGGATCCGCCAGCCCGTCCAGCAGCACCATG 3'

20 P10 (antisense)

# 5' CGCGCG<u>AAGCTT</u>TGTCAGCGCATGTTTTAATTGCAC 3'

in which the underlined palindromic sequences are the restriction sites for the enzymes BamHI (oligonucleotide P9) and HindIII (oligonucleotide P10). The PCR product was digested with the restriction enzymes BamHI and HindIII and then purified by electrophoresis on agarose gel.

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For the linker, the following partially complementary oligonucleotides were synthesized, and subsequently annealed to obtain a double strand DNA fragment with sticky ends:

P11 (sense)

5 5'AGCTTCGGGCGGTGGCGGTTCTGGTGGCGGTGGCCGGTGGCGGTTCT3'
P12 (antisense)

5CTAGAGAACCGCCACCGCCGCGAGCCACCGCCACCGCCACCGCCCGA3' in which the underlined bases are the sequences compatible with the restriction sites for the enzymes HindIII (oligonucleotide P11) and XbaI (oligonucleotide P12). The fragment resulting by PCR and the double strand linker sequence were inserted in the plasmid pRK7-Metron-F-1-His in place of the fragment EcoRI-XbaI by means of an EcoRI-BamHI adapter, to obtain the plasmid pRK7-Magic-F-1-His.

### Example 2b: Production of Magic Factor-1

Magic Factor-1 is produced on a small scale by transient transfection of BOSC cells analogously to what described for Metron Factor-1. Semi-purification is performed by adsorption on Sepharose-A beads conjugated with heparin followed by Western blot analysis using anti-poly-histidine antibodies (Fig. 5).

## Example 3: Biological activity (scattering) on epithelial cells.

The biological activity of recombinant HGF, NK2-HGF, Metron Factor-1 and Magic Factor-1 was tested by a "scatter" assay on MDCK epithelial cells. For this functional test, cells are plated at day 0 in 96-well plates (10<sup>3</sup> cells/well) in 90% DMEM - 10% bovine calf serum. At day 1 the medium is substituted with fresh medium buffered with 50 mM

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HEPES pH 7.4 and the supernatant containing the recombinant protein is added at different dilutions. At day 2 cells are washed with DPBS (Dulbecco's Phosphate Buffered Saline), fixed in 11% glutaraldehyde, stained with a Crystal-Violet solution and analysed by microscopy. The scattering activity is evaluated observing the morphology of the colonies, which are clustered in the negative control (non-stimulated cells or stimulated with supernatant containing no factors) whereas they are dispersed in the positive control (HGF-His). The morphology of the cells themselves also varies upon stimulation: in fact, as it can be observed in Fig. 6, cells stimulated with HGF-His and Metron Factor-1 have a more oblong, spindle-shaped form, characterised by protrusions of the cell membrane called pseudopodes. These morphological variations are the consequence of factor-induced activation of a genetic program involving the modification of a series of cellular parameters, such as digestion of cell matrix by specific proteases and increase in motility.

The Table summarises the results of different tests, obtained with factors HGF, NK2-HGF, Metron Factor-1 and Magic Factor-1 on MDCK cells. The scattering units reported indicate the maximum dilution of the conditioned supernatant containing the factor, at which motogenic activity could be observed. Values are normalised for the protein content determined by western blotting as described above (see Fig. 5). These data indicate that the hybrid factors Metron Factor-1 and Magic Factor-1 have a scattering activity approximately three magnitudes higher than that of the NK2-HGF-His truncated form and one magnitude higher than that of HGF-His parental factor.

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	HGF-his	NK2-his	Metron F-1	Magic Factor-1
Scatter units	$900 \pm 29$	6 ± 5	$5500 \pm 1532$	$7600 \pm 150$

Table. Scattering activity of factors HGF-His, NK2-HGF-His Metron Factor-1 measured on kidney epithelial cells (MDCK). The scattering units reported indicate the maximum dilution of the conditioned supernatant containing the factor, at which a motogenic activity can be observed. Values are normalised for the protein content determined by western blotting.

# Example 4a: Test for the evaluation of protection against programmed cell death (apoptosis).

One of the most characterised side effect of the chemotherapeutic drug cisplatin is the induction of programmed cell death (apoptosis) of epithelial cells of the proximal tubule, which leads to acute renal failure (ARF). Thus, a factor that protects against cisplatin-induced cytotoxicity is highly desirable. An in vitro functional test has been used, which allows to evaluate the percentage of cisplatin-treated apoptotic cells in the presence or in the absence of a survival factor. This system utilises a cell line (LOC) derived from epithelial cells of human kidney proximal tubule, immortalised by ectopic expression of SV40 large T antigen. For the functional test, cells are plated at day 0 in 96-well plates (10<sup>3</sup> cells/well) in 90% DMEM - 10% bovine calf serum. At day 1, the medium is substituted with medium containing 0.5% bovine calf serum buffered with 50 mM HEPES pH 7.4, which is added with different dilutions of the supernatant containing the recombinant factor. Cells are pre-incubated with these

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factors for 6 hours, and then further incubated in the presence of 10 µg/ml cisplatin. At day 2, cells are washed with DPBS and the percentage of apoptotic cells is evaluated by the TUNEL technique (Boehringer Mannheim). The same kind of tests can be performed using primary cultures of human epithelial cells of kidney proximal tubule (PTEC). These tests proved that Metron Factor-1 and Magic Factor-1 have protecting activity against cisplatin-induced programmed cell death.

# Example 4b: Protection against cisplatin-induced cytotoxicity by transient gene delivery of Metron Factor-1 and Magic Factor-1

The protective effect of Metron F-1 and Magic F-1 against cisplatin-induced cytotoxicity was further demonstrated by a transient gene delivery approach. Simian kidney epithelial cells (COS) were transfected with a control empty vector, an expression vector for Metron F-1, or an expression vector for Magic F-1. Following transfection, cells were treated for 16 hours with cisplatin (20 µg/ml) and the percentage of surviving cells in each transfection was determined. Cisplatin treatment was calibrated to cause the death of approximately 20% of the cells in the negative control. Ectopic expression of Metron F-1 or Magic F-1 increased the survival rate to about 92.3% and 94.0%, respectively.

# Example 5: Activation of the Met receptor by Metron Factor-1 and Magic Factor-1

The ability of Metron Factor-1 and Magic Factor-1 to activate the Met receptor was tested by analysing the ability of the recombinant factors to induce tyrosine phosphorylation of Met in human epithelial cells (A549). For this analysis, A549 cells at 90% confluence in a 100 mm petri

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dish were stimulated for 10 minutes with 1 ml of conditioned supernatant containing Metron Factor-1, Magic Factor-1 or no factor (as negative control) diluted 1:2.5 or 1:10 in DMEM. After stimulation, cells were washed in ice with PBS, lysated in 200 µl of lysis solution (1% Triton X-100, 5 mM EDTA, 150 mM NaCl, 20 mM Tris-HCl pH 7.4), added with a cocktail of protease inhibitors, immunoprecipitated for 2 hours at 4° C with 10 µl of Sepharose-A beads covalently conjugated with an anti-Met monoclonal antibody (Naldini, L. et al., 1991, EMBO J. 10: 2867-2878), washed 3 times in the same lysis solution, and heated at 90°C for 2 minutes to elute the absorbed proteins. These were separated by SDS-PAGE on a 8% BIS-acrylamide gel, transferred onto a membrane (Hybond-C; Amersham) and analysed by western blot. A mouse monoclonal antibody against phosphotyrosine (UBI) diluted 1:10000 was used as primary antibody and an anti-mouse IgG ovine antibody conjugated with peroxidase (Amersham) was used as secondary antibody. The secondary antibody was detected by ECL (Amersham) following the protocol provided by the manufacturer. This analysis revealed that Metron F-1 and Magic F-1 potently activate the Met receptor (Fig. 7).

# Example 6: Protection against chemotherapy-induced renal failure by Metron Factor-1 in vivo

Metron-F-1 was tested in a model of nephrotoxicity in Balb-c mice. The method used was substantially as described (Kawaida K et al., 1994, Hepatocyte growth factor prevents acute renal failure and accelerates renal regeneration in mice, Proc. Natl. Acad. Sci. 91:4357-4361). Briefly, renal failure was induced in male Balb-c mice weighing 20-25 g by an i.v.

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injection of 7.5 mg/kg of HgCl<sub>2</sub> (7 animals/group). Renal damage was assessed by analysis of Blood Urea Nitrogen (BUN) and by histological evaluation, 72 h after HgCl<sub>2</sub> injection. Metron-F-1 was dissolved in 0.2 M NaCl, containing 0.01% Tween 80 and 0.25% human serum albumin and administered i.v. (100 μg/kg in a posological volume of 6.6 ml/kg) 0.5 h before and 6, 12, 24, 36 and 48 h after HgCl<sub>2</sub> injection. Controls animals were treated with the same amount of vehicle according to the same scheme.

Metron-F-1 significantly prevented the onset of acute renal failure induced by HgCl<sub>2</sub>, evaluated in terms of BUN (figure 8). BUN values were closely paralleled by the histological findings, scored by an independent investigator.

In the following sequence listing:

SEQ. ID. NO. 1: Magic F-1 DNA coding sequence;

SEQ. ID. NO. 2: Magic F-1 amino acid sequence;

SEQ. ID. NO. 3: Metron F-1 DNA coding sequence;

SEQ. ID. NO. 4: Metron F-1 amino acid sequence.

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## **CLAIMS**

1. Recombinant proteins comprising two superdomains, separated by a spacer sequence (linker), obtained combining the HL and K1-K4 domains of HGF and/or MSP α chains, according to general formula (I):

$$[A] - B - [C] - (D)y$$
 (I)

in which

[A] corresponds to the sequence (LS)<sub>m</sub>-HL-K1-(K2)<sub>n</sub>-(K3)<sub>o</sub>-(K4)<sub>p</sub> wherein (the numbering of the following amino acids refers to the HGF and MSP sequences as reported in Fig. 1 and 2, respectively):

LS is an amino acid sequence corresponding to residues 1-31 of HGF or 1-18 of MSP;

HL is an amino acid sequence starting between residues 32-70 of HGF  $\alpha$  chain and ending between residues 96-127 of the identical chain; or it is an amino acid sequence starting between residues 19-56 of MSP  $\alpha$  chain and ending between residues 78-109 of the identical chain;

K1 is an amino acid sequence starting between residues 97-128 of HGF  $\alpha$  chain and ending between residues 201-205 of the identical chain; or it is an amino acid sequence starting between residues 79-110 of MSP  $\alpha$  chain and ending between residues 186-190 of the identical chain;

K2 is an amino acid sequence starting between residues 202-206 of HGF  $\alpha$  chain and ending between residues 283-299 of the identical chain; or it is an amino acid sequence starting between residues 187-191 of MSP  $\alpha$  chain and ending between residues 268-282 of the identical chain;

K3 is an amino acid sequence starting between residues 284-300 of HGF α chain and ending between residues 378-385 of the identical chain; or it is

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an amino acid sequence starting between residues 269-283 of MSP a chain and ending between residues 361-369 of the identical chain;

K4 is an amino acid sequence starting between residues 379-386 of HGF  $\alpha$ chain and ending between residues 464-487 of the identical chain; or it is an amino acid sequence starting between residues 362-370 of MSP a chain and ending between residues 448-481 of the identical chain;

m, n, o, p are 0 or 1;

the sum n + o + p is an integer from 1 to 3 or 0, with the proviso that n  $\geq 0 \geq p$ ;

B is the sequence  $[(X)_q Y]_{\tau}$ , wherein X = Gly and Y = Ser, or Cys, or Met, or Ala;

q is an integer from 2 to 8;

r is an integer from 1 to 9;

[C] corresponds to the sequence HL-K1-(K2)<sub>s</sub>-(K3)<sub>t</sub>-(K4)<sub>n</sub>

wherein HL, K1-K4 are as defined above,

s, t, u are 0 or 1; the sum s + t + u is an integer from 1 to 3 or 0, with the proviso that  $s \ge t \ge u$ ;

D is the sequence W-Z, wherein W is a conventional proteolytic site, Z is any tag sequence useful for the purification and detection of the protein; y is 0 or 1.

Recombinant proteins according to claims 1-2, in which the HL domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 32 to 127, or a sequence of MPS a chain ranging from amino acids 19 to 98; the K1 domain is a sequence of HGF \alpha chain ranging from amino acids 128 to 203, or a sequence of MPS  $\alpha$  chain ranging from amino acids 99 to 188; the K2 domain is a sequence of HGF \alpha chain ranging from amino acids 204 to 294, . 13- 3- V :

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or a sequence of MPS  $\alpha$  chain ranging from amino acids 189 to 274; the K3 domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 286 to 383, or a sequence of MPS  $\alpha$  chain ranging from amino acids 275 to 367; the K4 domain is a sequence of HGF  $\alpha$  chain ranging from amino acids 384 to 487, or a sequence of MPS  $\alpha$  chain ranging from amino acids 368 to 477.

- 3. Recombinant proteins according to claims 1-2 of formula (II):
- LS<sub>MSP</sub>-HL<sub>MSP</sub>-K1<sub>MSP</sub>-K2<sub>MSP</sub>-L-HL<sub>HGF</sub>-K1<sub>HGF</sub>-K2<sub>HGF</sub>-D (II) in which LS<sub>MSP</sub> is the sequence 1-18 of MSP, HL<sub>MSP</sub> is the sequence 19-56 of MSP, K1<sub>MSP</sub> is the sequence 99-188 of MSP, K2<sub>MSP</sub> is the sequence 189-274 of MSP, HL<sub>HGF</sub> is the sequence 32-127 of HGF, K1<sub>HGF</sub> is the sequence 128-203 of HGF, K2<sub>HGF</sub> is the sequence 204-294 of HGF, L is the sequence (Gly<sub>4</sub>Ser)<sub>3</sub>, D is the sequence Asp<sub>4</sub>-Lys-His<sub>6</sub>.
- 4. Recombinant proteins according to claims 1-2 of formula (III):

  LS<sub>HGF</sub>-HL<sub>HGF</sub>-K1<sub>HGF</sub>-K2<sub>HGF</sub>-L-HL<sub>HGF</sub>-K1<sub>HGF</sub>-K2<sub>HGF</sub>-D (III)

  in which HL<sub>HGF</sub>, K1<sub>HGF</sub>, K2<sub>HGF</sub>, L and D are as defined in claim 4, LS<sub>HGF</sub> is
- 5. Nucleotide sequences encoding for the recombinant proteins of claims 1-5.
- 6. Expression vectors comprising the nucleotide sequences of claim 5.
- 7. Prokaryotic or eukaryotic host cell transformed with the expression vector of claim 6.
  - 8. Process for preparing the recombinant proteins of claims I-4, which comprises the following steps:
  - a) construction of DNA encoding the desired protein;
- 25 b) insertion of DNA in an expression vector;

the sequence 1-31 of HGF.

c) transformation of a host cell with recombinant DNA (rDNA);

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- d) culture of the transformed host cell so as to express the recombinant protein;
- e) extraction and purification of the produced recombinant protein.
- 9. Process according to claim 8, wherein the host cell is kidney epithelial BOSC cell or SF9 insect cell.
- 10. Recombinant proteins of claims 1-4 for use as therapeutic agents.
- 11. Use of recombinant proteins of claims 1-4 in the manufacture of a medicament for the prevention or treatment of chemotherapeutic-induced toxicity.
- 12. Use according to claim 9, wherein the chemotherapeutic-induced toxicity is myelotoxicity, kidney toxicity, neurotoxicity, mucotoxicity and hepatotoxicity.
- 13. Pharmaceutical compositions containing an effective amount of the recombinant proteins of claims 1-4, in combination with pharmacologically acceptable excipients.

#### SEQUENCE LISTING

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(1) GENERAL I	NFORMATION:
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- (i) APPLICANT:
  - (A) NAME: DOMPE' S.p.A.
  - (B) STREET: Via Campo di Pile
  - (C) CITY: L'AQUILA (E) COUNTRY: ITALY
  - (F) POSTAL CODE (ZIP): 67100
- (ii) TITLE OF INVENTION: RECOMBINANT PROTEINS DERIVED FROM HGF AND MSP
- (iii) NUMBER OF SEQUENCES: 4
- (iv) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Floppy disk
  - (B) COMPUTER: IBM PC compatible
  - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
  - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)
- (2) INFORMATION FOR SEQ ID NO: 1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1725 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (iii) HYPOTHETICAL: NO
    - (iv) ANTI-SENSE: NO
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

ATGTGGGTGA C	CAAACTCCT	GCCAGCCCTG	CTGCTGCAGC	ATGTCCTCCT	GCATCTCCTC	60
CTGCTCCCCA T	CCCCATCCC	CTATGCAGAG	GGACAAAGGA	AAAGAAGAAA	TACAATTCAT	120
GAATTCAAAA A	AATCAGCAAA	GACTACCCTA	ATCAAAATAG	ATCCAGCACT	GAAGATAAAA	180
ACCAAAAAAG I	rgaatactgc	AGACCAATGT	GCTAATAGAT	GTACTAGGAA	TAAAGGACIT	240
CCATTCACTT 6	GCAAGGCTTT	TGTTTTTGAT	AAAGCAAGAA	AACAATGCCT	CTGGTTCCCC	300
TTCAATAGCA	TGTCAAGTGG	AGTGAAAAAA	GAATTTGGCC	ATGAATTTGA	CCTCTATGAA	360
AACAAAGACT A	ACATTAGAAA	CTGCATCATT	GGTAAAGGAC	GCAGCTACAA	GGGAACAGTA	420
TCTATCACTA	AGAGTGGCAT	CAAATGTCAG	CCCTGGAGTT	CCATGATACC	ACACGAACAC	480
AGCTATCGGG (	GTAAAGACCT	ACAGGAAAAC	TACTGTCGAA	ATCCTCGAGG	GGAAGAAGGG	540
GGACCCTGGT	GTTTCACAAG	CAATCCAGAG	GTACGCTACG	AAGTCTGTGA	CATTCCTCAG	600

rgttcagaag	TTGAATGCAT	GACCTGCAAT	GGGGAGAGTT	ATCGAGGTCT	CATGGATCAT	660
ACAGAATCAG	GCAAGATTTG	TCAGCGCTGG	GATCATCAGA	CACCACACCG	GCACAAATTC	720
TTGCCTGAAA	GATATCCCGA	CAAGGCTTT	GATGATAATT	ATTGCCGCAA	TCCCGATGGC	780
CAGCCGAGGC	CATGGTGCTA	TACTCTTGAC	CCTCACACCC	GCTGGGAGTA	CTGTGCAATT	840
AAAACATGCG	CTGACAAAGC	TTCGGGCGGT	GGCGGTTCTG	GTGGCGGTGG	CTCCGGCGGT	900
GGCGGTTCTC	TAGAGGGACA	AAGGAAAAGA	AGAAATACAA	TTCATGAATT	CAAAAAATCA	960
GCAAAGACTA	CCCTAATCAA	AATAGATCCA	GCACTGAAGA	TAAAAACCAA	AAAAGTGAAT	1020
ACTGCAGACC	AATGTGCTAA	TAGATGTACT	AGGAATAAAG	GACTTCCATT	CACTTGCAAG	1080
GCTTTTGTTT	TTGATAAAGC	AAGAAAACAA	TGCCTCTGGT	TCCCCTTCAA	TAGCATGTCA	1140
AGTGGAGTGA	AAAAAGAATT	TGGCCATGAA	TTTGACCTCT	ATGAAAACAA	AGACTACATT	1200
AGAAACTGCA	TCATTGGTAA	AGGACGCAGC	TACAAGGGAA	CAGTATCTAT	CACTAAGAGT	1260
GGCATCAAAT	GTCAGCCCTG	GAGTTCCATG	ATACCACACG	AACACAGCTA	TCGGGGTAAA	1320
GACCTACAGG	AAAACTACTG	TCGAAATCCT	CGAGGGGAAG	AAGGGGGACC	CIGGIGITIC	1380
ACAAGCAATC	CAGAGGTACG	CTACGAAGTO	TGTGACATTC	CTCAGTGTTC	AGAAGTTGAA	1440
TGCATGACCI	GCAATGGGGA	GAGTTATCGA	GGTCTCATGG	ATCATACAGA	ATCAGGCAAG	1500
ATTTGTCAGO	C GCTGGGATCA	TCAGACACC	A CACCGGCACA	AATTCTTGC	C TGAAAGATAT	1560
CCCGACAAGG	GCTTTGATG	TAATTATTG	C CGCAATCCCG	aTGGCCAGC	C GAGGCCATGG	1620
TGCTATACTO	TTGACCCTC	A CACCCGCTG	GAGTACTGT	CAATTAAAA	C ATGCGCTGAC	: 1680
AAAGCTGAC	ACGACGACA	A ACACCACCA	C CACCACCAC	CACTAG		1725

- (2) INFORMATION FOR SEQ ID NO: 2:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 574 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Trp Val Thr Lys Leu Leu Pro Ala Leu Leu Leu Gln His Val Leu 1 5 10 15

Leu His Leu Leu Leu Leu Pro Ile Ala Ile Pro Tyr Ala Glu Gly Gln 20 25 30

Arg Lys Arg Arg Asn Thr Ile His Glu Phe Lys Lys Ser Ala Lys Thr 35 40 45

Thr Leu Ile Lys Ile Asp Pro Ala Leu Lys Ile Lys Thr Lys Lys Val 50 55 60

Asn Thr Ala Asp Gln Cys Ala Asn Arg Cys Thr Arg Asn Lys Gly Leu 65 70 75 80

Pro Phe Thr Cys Lys Ala Phe Val Phe Asp Lys Ala Arg Lys Gln Cys 85 90 95

Leu Trp Phe Pro Phe Asn Ser Met Ser Ser Gly Val Lys Lys Glu Phe 100 105 110

Gly His Glu Phe Asp Leu Tyr Glu Asn Lys Asp Tyr Ile Arg Asn Cys 115 120 125

Ile Ile Gly Lys Gly Arg Ser Tyr Lys Gly Thr Val Ser Ile Thr Lys 130 135 140

Ser Gly Ile Lys Cys Gln Pro Trp Ser Ser Met Ile Pro His Glu His 145 150 155 160

Ser Tyr Arg Gly Lys Asp Leu Gln Glu Asn Tyr Cys Arg Asn Pro Arg 165 170 175

Gly Glu Glu Gly Gly Pro Trp Cys Phe Thr Ser Asn Pro Glu Val Arg 180 185 190

Tyr Glu Val Cys Asp Ile Pro Gln Cys Ser Glu Val Glu Cys Met Thr 195 200 205

Cys Asn Gly Glu Ser Tyr Arg Gly Leu Met Asp His Thr Glu Ser Gly 210 215 220

Lys Ile Cys Gln Arg Trp Asp His Gln Thr Pro His Arg His Lys Phe 230 Leu Pro Glu Arg Tyr Pro Asp Lys Gly Phe Asp Asp Asn Tyr Cys Arg 250 Asn Pro Asp Gly Gln Pro Arg Pro Trp Cys Tyr Thr Leu Asp Pro His 265 260 Thr Arg Trp Glu Tyr Cys Ala Ile Lys Thr Cys Ala Asp Lys Ala Ser 280 Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser Leu 295 290 300 Glu Gly Gln Arg Lys Arg Arg Asn Thr Ile His Glu Phe Lys Lys Ser 310 315 Ala Lys Thr Thr Leu Ile Lys Ile Asp Pro Ala Leu Lys Ile Lys Thr 325 330 Lys Lys Val Asn Thr Ala Asp Gln Cys Ala Asn Arg Cys Thr Arg Asn 340 345 Lys Gly Leu Pro Phe Thr Cys Lys Ala Phe Val Phe Asp Lys Ala Arg 360 365 Lys Gln Cys Leu Trp Phe Pro Phe Asn Ser Met Ser Ser Gly Val Lys 375 Lys Glu Phe Gly His Glu Phe Asp Leu Tyr Glu Asn Lys Asp Tyr Ile 390 385 395 Arg Asn Cys Ile Ile Gly Lys Gly Arg Ser Tyr Lys Gly Thr Val Ser 410 Ile Thr Lys Ser Gly Ile Lys Cys Gln Pro Trp Ser Ser Met Ile Pro 425 430 His Glu His Ser Tyr Arg Gly Lys Asp Leu Gln Glu Asn Tyr Cys Arg 435 Asn Pro Arg Gly Glu Glu Gly Gly Pro Trp Cys Phe Thr Ser Asn Pro Glu Val Arg Tyr Glu Val Cys Asp Ile Pro Gln Cys Ser Glu Val Glu 465 470 480 Cys Met Thr Cys Asn Gly Glu Ser Tyr Arg Gly Leu Met Asp His Thr 490 Glu Ser Gly Lys Ile Cys Gln Arg Trp Asp His Gln Thr Pro His Arg 500 505 510 His Lys Phe Leu Pro Glu Arg Tyr Pro Asp Lys Gly Phe Asp Asp Asn 515 520

Tyr	Cys	Arg	Asn	Pro	Asp	Gly	${\tt Gln}$	Pro	Arg	${\tt Pro}$	Trp	Cys	Tyr	Thr	Leu
-	530	-				535					540				

Asp Pro His Thr Arg Trp Glu Tyr Cys Ala Ile Lys Thr Cys Ala Asp 545 550 555 560

Lys Ala Asp Asp Asp Lys His His His His His His His 565 570

#### (2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1692 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

ATGGGGTGGC	PCCCACTCCT	GCTGCTTCTG	ACTCAATGCT	TAGGGGTCCC	TGGGCAGCGC	60
TCGCCATTGA A	ATGACTTCCA	AGTGCTCCGG	GGCACAGAGC	TACAGCACCT	GCTACATGCG	120
GTGGTGCCCG (	GGCCTTGGCA	GGAGGATGTG	GCAGATGCTG	AAGAGTGTGC	TGGTCGCTGT	180
GGGCCCTTAA '	TGGACTGCCG	GGCCTTCCAC	TACAACGTGA	GCAGCCATGG	TTGCCAACTG	240
CTGCCATGGA (	CTCAACACTC	GCCCCACACG	AGGCTGCGGC	GTTCTGGGCG	CTGTGACCTC	300
TTCCAGAAGA	AAGACTACGT	ACGGACCTGC	ATCATGAACA	ATGGGGTTGG	GTACCGGGGC	360
ACCATGGCCA	CGACCGTGGG	TGGCCTGCCC	TGCCAGGCTT	GGAGCCACAA	GTTCCCGAAT	420
GATCACAAGT .	ACACGCCCAC	TCTCCGGAAT	GGCCTGGAAG	AGAACTTCTG	CCGTAACCCT	480
GATGGCGACC	CCGGAGGTCC	TTGGTGCTAC	ACAACAGACC	CIGCIGIGCG	CTTCCAGAGC	540
TGCGGCATCA	AATCCTGCCG	GGAGGCCGCG	TGTGTCTGGT	GCAATGGCGA	GGAATACCGC	600
GGCGCGGTAG	ACCGCACGGA	GTCAGGGCGC	GAGTGCCAGC	GCTGGGATCT	TCAGCACCCG	660
CACCAGCACC	CCTTCGAGCC	GGGCAAGTTC	CTCGACCAAG	GTCTGGACGA	CAACTATTGC	720
CGGAATCCTG	ACGGCTCCGA	GCGGCCATGG	TGCTACACTA	CGGATCCGCA	GATCGAGCGA	780
GAGTTCTGTG	ACCTCCCCCG	CTGCGGGTCC	GAGGCACAGC	CCCGCCTCGA	GGGCGGTGGC	840
GGTTCTGGTG	GCGGTGGCTC	CGGCGGTGGC	GGTTCTCTAG	AGGGACAAAG	GAAAAGAAGA	900
AATACAATTC	ATGAATTCAA	AAAATCAGCA	AAGACTACCO	TAATCAAAAT	AGATCCAGCA	960

AATAAAGGAC TTCCATTCAC TTGCAAGGCT TTTGTTTTTG ATAAAGCAAG AAAACAATGC 1080
CTCTGGTTCC CCTTCAATAG CATGTCAAGGCT GGAGTGAAAA AAGAATTTGG CCATGAATTT 1140
GACCTCTATG AAAACAAAGA CTACATTAGA AACTGCATCA TTGGTAAAGG ACGCAGCTAC 1200
AAGGGAACAG TATCTATCAC TAAGAGTGGC ATCAAATGTC AGCCCTGGAG TTCCATGATA 1260
CCACACGGAAC ACAGCTATCG GGGTAAAGAC CTACAGGAAA ACTACTGTCG AAATCCTCGA 1320
GGGGAAGAAG GGGGACCCTG GTGTTCACA AGCAATCCAG AGGTACGCTA CGAAGTCTGT 1380
GACATTCCTC AGTGTTCAGA AGTTGAATGC ATGACCTGCA ATGGGGAAGA TTATCGAGGT 1440
CTCATGGATC ATACAGAATC AGGCAAGATT TGTCAGCGCT GGGATCATCA GACACCACAC 1500
CGGCACAAAT TCTTGCCTGA AAGATATCCC GACAAGGGCT TTGATGATAA TTATTGCCGC 1560
AATCCCGATG GCCAGCCGAG GCCATGGTGC TATACTCTTG ACCCTCACAC CCGCTGGGAG 1620
TACTGTGCAA TTAAAACATG CGCTGACAAA GCTGACGACG ACGACAAACA CCACCACCA 1680
CACCACCACT AG

#### (2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 563 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Gly Trp Leu Pro Leu Leu Leu Leu Leu Thr Gln Cys Leu Gly Val 1 5 10 15

Pro Gly Gln Arg Ser Pro Leu Asn Asp Phe Gln Val Leu Arg Gly Thr 20 25 30

Glu Leu Gln His Leu Leu His Ala Val Val Pro Gly Pro Trp Gln Glu 35 40 45

Asp Val Ala Asp Ala Glu Glu Cys Ala Gly Arg Cys Gly Pro Leu Met 50 55 60

Asp Cys Arg Ala Phe His Tyr Asn Val Ser Ser His Gly Cys Gln Leu 65 70 75 80

Leu Pro Trp Thr Gln His Ser Pro His Thr Arg Leu Arg Arg Ser Gly 85 90 95

Arg Cys Asp Leu Phe Gln Lys Lys Asp Tyr Val Arg Thr Cys Ile Met 100 105 110

Asn Asn Gly Val Gly Tyr Arg Gly Thr Met Ala Thr Thr Val Gly Gly 115 120 125

Leu Pro Cys Gln Ala Trp Ser His Lys Phe Pro Asn Asp His Lys Tyr 130 135 140

Thr Pro Thr Leu Arg Asn Gly Leu Glu Glu Asn Phe Cys Arg Asn Pro 145 150 155 160

Asp Gly Asp Pro Gly Gly Pro Trp Cys Tyr Thr Thr Asp Pro Ala Val 165 170 175

Arg Phe Gln Ser Cys Gly Ile Lys Ser Cys Arg Glu Ala Ala Cys Val 180 185 190

Trp Cys Asn Gly Glu Glu Tyr Arg Gly Ala Val Asp Arg Thr Glu Ser 195 200 205

Gly Arg Glu Cys Gln Arg Trp Asp Leu Gln His Pro His Gln His Pro 210 215 220

Phe Glu Pro Gly Lys Phe Leu Asp Gln Gly Leu Asp Asp Asn Tyr Cys 225 230 235 240

Arg Asn Pro Asp Gly Ser Glu Arg Pro Trp Cys Tyr Thr Thr Asp Pro 245 250 255

Gln Ile Glu Arg Glu Phe Cys Asp Leu Pro Arg Cys Gly Ser Glu Ala 260 265 270

Gln Pro Arg Leu Glu Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly 275 280 285

Gly Gly Ser Leu Glu Gly Gln Arg Lys Arg Arg Asn Thr Ile His 290 295 300

Glu Phe Lys Lys Ser Ala Lys Thr Thr Leu Ile Lys Ile Asp Pro Ala 305 310 315 320

Leu Lys Ile Lys Thr Lys Lys Val Asn Thr Ala Asp Gln Cys Ala Asn 325 330 335

Arg Cys Thr Arg Asn Lys Gly Leu Pro Phe Thr Cys Lys Ala Phe Val 340 345 350

Phe Asp Lys Ala Arg Lys Gln Cys Leu Trp Phe Pro Phe Asn Ser Met 355 360 365

Ser Ser Gly Val Lys Lys Glu Phe Gly His Glu Phe Asp Leu Tyr Glu 370 375 380 Asn Lys Asp Tyr Ile Arg Asn Cys Ile Ile Gly Lys Gly Arg Ser Tyr 385 390 395 400

Lys Gly Thr Val Ser Ile Thr Lys Ser Gly Ile Lys Cys Gln Pro Trp 405 410 415

Ser Ser Met Ile Pro His Glu His Ser Tyr Arg Gly Lys Asp Leu Gln 420 425 430

Glu Asn Tyr Cys Arg Asn Pro Arg Gly Glu Glu Gly Gly Pro Trp Cys
435
440
445

Phe Thr Ser Asn Pro Glu Val Arg Tyr Glu Val Cys Asp Ile Pro Gln 450 455 460

Cys Ser Glu Val Glu Cys Met Thr Cys Asn Gly Glu Ser Tyr Arg Gly 465 470 475 480

Leu Met Asp His Thr Glu Ser Gly Lys Ile Cys Gln Arg Trp Asp His 485 490 495

Gln Thr Pro His Arg His Lys Phe Leu Pro Glu Arg Tyr Pro Asp Lys 500 505 510

Gly Phe Asp Asp Asn Tyr Cys Arg Asn Pro Asp Gly Gln Pro Arg Pro 515 520 525

Trp Cys Tyr Thr Leu Asp Pro His Thr Arg Trp Glu Tyr Cys Ala Ile 530 535 540

Lys Thr Cys Ala Asp Lys Ala Asp Asp Asp Asp Lys His His His 545 550 555 560

His His His

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#### FIG 1a

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CA) 4	UAC! W	V.	CIG T	E I	i Gad	ADD L	و ع	a A	gga L				CG1. H	ACA! V	gga L		СС1. Н	agai L	L
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CT	GCT	CCC	CAT	CGC	CAT	CCC	CTA	TGC	AGA	GGG	ACA	AAG	GAA	AAG.	AAG	AAA	TAC.	TAA	TCA
JA(	CGA	 GGG	-+- GTA									TTC				TTT.	ATG	TTA	AGTA
	L	Ъ	I	A	I	P	Y	A	Ε	G	Q	R	K	R	R	N	T	I	Н
			+-							+			-+-			+			AAA!
T'	TAA	GTT	TTT	TAG	TCG	TTT	CTG	ATG	GGA	TTA	.GTT	TTA	TCT	AGG	TCG	TGA	CTT	CTA	TTT
2	F	K	K	S	A.	K	Ţ	Ţ	L	I	K	I	D	D,	A	L	K	Ι	K
			-+-			÷				+			-+-			+			ACT:
•	K	K	V	N	T	A	D	Q	С	A	N	R	С	T	R	И	K	G	L
:c̄	ATT	CAC	TTG	CAA	.GGC	TTT	TGT	TTT	TGA	TAA	AGC	:AAG	AAA	ACA	ATG	CCT	CTG	GTT	CCC
G	TAA	GTG	AAC	GTT	CCG	AAA	ACA	AAA	ACT	ATT	TCC	TTC	TTT	TGT	TAC	GGA	.GAC	CAA	.GGG
?	F	T	С	K	A	F	V	Ê	D	K	A	R	K	Q	С	L	W	Ε	P
						TGG	AGT	'GAA	<u>מממ</u>	AGP	LATT	TGG	CCA	TGE	דיתב	'TGA	CCT	רייים א	шСл
T	CAA	TAG	CAT	GTC	AAG														.1GA
			-+-			+		.CTT				LACC	-+-			÷			
λA			-+-			+				TCI			-+-			÷			
AA	GTT N	ATC	-+- GTA M	.CAG	TTC S	ACC	TCA V	K	K	TCI E	TAA F	G	GGT	ACT E	TAA	ACT D	'GGA L	GAT Y	ACT
A.	GTT. N CAA	ATC S AGA	GTA M .CTA	CAG	TTC	ACC G	TCA V	K CAT	TTT	TCI E	TAA F	G AAGG	H ACG	ACT E	TAP F	ACAA	'GGA L	GAT Y	ACT E E AGT
IA.	GTT N CAA GTT	ATC S AGA TCT	GTA M .CTA	CAG	TTC	ACC G	V CTG	K CAT	TTTT  K 'CAI	TCI E TGG	TAA F	G AAGG TTCC	H ACG	ACT E CAG	TAP F	ACAA	L L LGGG	GAT Y	ACT E E AGT
IAI	GTT N CAA GTT K	ATC S AGA TCT D	GTA  M  CTA  GAT  Y	CAG S CAT CAT I	TAG ATC	G G G AAA TTT N GCAT	V CTG GAC	K CAT GTA I	CAT  CAT  GTCF	TTCT E TTGG ACC	TAA	G AAGO TTCO G GGAO	GGT H SACG TGC R	ACT E GCAG GTC	TAA F GCTA GAI	ACAA	GGA L L CCCC G	GAT Y AAC TTG	ACT E AGT TCA V
AA TT	GTT N CAA GTT K	ATC S AGA TCT D	GTA  M  CTA  GAT  Y	CAG S CAT CAT I	TAG ATC	G G G AAA TTT N GCAT	V CTG GAC	K CAT GTA I	CAT  CAT  GTCF	TTCT E TTGG ACC	TAA	G AAGO TTCO G GGAO	GGT H SACG TGC R	ACT E GCAG GTC	TAA F GCTA GAI	ACAA	GGA L L CCCC G	GAT Y AAC TTG	ACT E AGT TCA

	(continued) 2/19	
	AGCTATCGGGGTAAAGACCTACAGGAAAACTACTGTCGAAATCCTCGAGGGGAAGAAGGG	
481	TCGATAGCCCCATTTCTGGATGTCCTTTTGATGACAGCTTTAGGAGCTCCCCTTCTTCCC	540
161	SYRGKDLQENYCRNPRGEEG	180
541	GGACCCTGGTGTTTCACAAGCAATCCAGAGGTACGCTACGAAGTCTGTGACATTCCTCAG	600
181	G P W C F T S N P E V R Y E V C D I P Q	200
601	TGTTCAGAAGTTGAATGCATGACCTGCAATGGGGAGAGTTATCGAGGTCTCATGGATCAT ACAAGTCTTCAACTTACGTACTGGACGTTACCCCTCTCAATAGCTCCAGAGTACCTAGTA	660
201	C S E V E C M T C N G E S Y R G L M D H	220
661	ACAGAATCAGGCAAGATTTGTCAGCGCTGGGATCATCAGACACCACACCGGCACAAATTC TGTCTTAGTCCGTTCTAAACAGTCGCGACCCTAGTAGTCTGTGGTGTGGCCGTGTTTAAG	720
221	T E S G K I C Q R W D H Q T P H R H K F	240
721	TTGCCTGAAAGATATCCCGACAAGGGCTTTGATGATAATTATTGCCGCAATCCCGATGGC+ AACGGACTTTCTATAGGGCTGTTCCCGAAACTACTATTAATAACGGCGTTAGGGCTACCG	780
241	L P E R Y P D K G F D D N Y C R N P D G	260
781	CAGCCGAGGCCATGGTGCTATACTCTTGACCCTCACACCCGCTGGGAGTACTGTGCAATT	840
261	Q P R P W C Y T L D P H T R W E Y C A I	280
841	AAAACATGCGCTGACAATACTATGAATGACACTGATGTTCCTTTGGAAACAACTGAATGC	900
281	K T C A D N T M N D T D V P L E T T E C	300
	ATCCAAGGTCAAGGAGAAGGCTACAGGGGCACTGTCAATACCATTTGGAATGGAATTCCA	
901	TAGGTTCCAGTTCCTCTCCGATGTCCCCGTGACAGTTATGGTAAACCTTACCTTAAGGT	960
301	I Q G Q G E G Y R G T V N T I W N G I P	320
961	TGTCAGCGTTGGGATTCTCAGTATCCTCACGAGCATGACATGACTCCTGAAAATTTCAAG ACAGTCGCAACCCTAAGAGTCATAGGAGTGCTCGTACTGTACTGAGGACTTTTAAAGTTC	1020
321	CQRWDSQYPHEHDMTPENFK	340
1021	TGCAAGGACCTACGAGAAAATTACTGCCGAAATCCAGATGGGTCTGAATCACCCTGGTGT ACGTTCCTGGATGCTCTTTTAATGACGGCTTTAGGTCTACCCAGACTTAGTGGGACCACA	1080
341	C K D L R E N Y C R N P D G S E S P W C	360
1081	TTTACCACTGATCCAAACATCCGAGTTGGCTACTGCTCCCAAATTCCAAACTGTGATATG AAATGGTGACTAGGTTTGTAGGCTCAACCGATGACGAGGGTTTAAGGTTTGACACTATAC	1140
361		380
	(continued)	١

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1141	TCACATGGACAAGATTGTTATCGTGGGAATGGCAAAAATTATATGGGCAACTTATCCCAA	1200
381	SHGQDCYRGNGKNYMGNLSQ	400
1201	ACAAGATCTGGACTAACATGTTCAATGTGGGACAAGAACATGGAAGACTTACATCGTCAT TGTTCTAGACCTGATTGTACAAGTTACACCCTGTTCTTGTACCTTCTGAATGTAGCAGTA	1260
401	TRSGLTCSMWDKNMEDLHRH	420
1261	ATCTTCTGGGAACCAGATGCAAGTAAGCTGAATGAGAATTACTGCCGAAATCCAGATGAT TAGAAGACCCTTGGTCTACGTTCATTCGACTTACTCTTAATGACGGCTTTAGGTCTACTA	1320
421	I F W E P D A S K L N E N Y C R N P D D	440
1321	GACGCTCATGGACCCTGGTGCTACACGGGAAATCCACTCATTCCTTGGGATTATTGCCCT CTGCGAGTACCTGGGACCACGATGTGCCCTTTAGGTGAGTAAGGAACCCTAATAACGGGA	1380
441	D A H G P W C Y T G N P L I P W D Y C P	460
1381	ATTTCTCGTTGTGAAGGTGATACCACACCTACAATAGTCAATTTAGACCATCCCGTAATA TAAAGAGCAACACTTCCACTATGGTGTGGATGTTATCAGTTAAATCTGGTAGGGCATTAT	1440
461	I S R C E G D T T P T I V N L D H P V I	480
1441	TCTTGTGCCAAAACGAAACAATTGCGAGTTGTAAATGGGATTCCAACACGAACAAACA	1500
481	S C A K T K Q L R V V N G I P T R T N I	500
1501	GGATGGATGGTTAGTTTGAGATACAGAAATAAACATATCTGCGGAGGATCATTGATAAAG CCTACCTACCAATCAAACTCTATGTCTTTATTTGTATAGACGCCTCCTAGTAACTATTTC	1560
501	G W M V S L R Y R N K H I C G G S L I K	520
1561	GAGAGTTGGGTTCTTACTGCACGACAGTGTTTCCCTTCTCGAGACTTGAAAGATTATGAA CTCTCAACCCAAGAATGACGTGCTGTCACAAAGGGAAGAGCTCTGAACTTTCTAATACTT	1620
521	E S W V L T A R Q C F P S R D L K D Y E	540
1621	GCTTGGCTTGGAATTCATGATGTCCACGGAAGAGGGAGATGAGAAATGCAAACAGGTTCTC CGAACCGAAC	1680
541	A W L G I H D V H G R G D E K C K Q V L	560
1681	AATSTTTCCCAGCTGGTATATGGCCCTGAAGGATCAGATCTGGTTTTAATGAAGCTTGCC TTACAAAGGGTCGACCATATACCGGGACTTCCTAGTCTAGACCAAAATTACTTCGAACGG	1740
561	N V S Q L V Y G P E G S D L V L M K L A	580
1741	AGGCCTGCTGTCCTGGATGATTTTGTTAGTACGATTGATT	1800
581	RPAVLDDFVSTIDLPNYGCT (continued)	600

# (continued)

	AT	TCC	TGA	AAA	GAC	CAGʻ	TTĢ	CAG'	rgt'	rta:	rgg	CTG	GGG	CTA	CAC	TGG.	TTA	GAT	CAA	CTAT	1860
1801	TA	AGG.	ACT	TTT	CTG	GTC	AAC	GTC	ACA	LTAP	ACC	GAC	ccc	GAT	GTG.	ACC	TAA	CTA	GTT	GATA	2000
601	I	P	Ε	K	Т	S	С	S	V	Y	G	W	G	Y	Т	G	L	Ξ	N	Y	620
1861											<del>+</del>		_ <del></del> -	-+-						GCAT	1920
3	CT	ACC	GGA	TAA	TGC	TCA	CCG	TGT.	AGA	GAT:	AŢA	ŢŢA	ccc	TTT	ACT.	CTT	ŢAC	GŢC	GGT	CGTA	
621	D.	G	L	L	R	V	A	н	L	Y	I	М	G	И	Ε	K	С	S	Q	H	640
1921		- · · ·					÷				<del></del>									TGGA	1980
	GT	AGC	TCC	CTT	CCA	CTG	AGA	CTT	ACT	CAG	ACT	TTA	TAC	ACG	ACC	CCG	ACT	TTT	CTA	ACCT	
641	Н	R	G	K	V	T	L	N	Ε	S	Ε	I	С	A	G	Ą	Ε	K	Ξ	G	660
1981																				GAGA	2040
	AG	TCC	TGG	TAC	ACI	CCC	CCT	'AAT	'ACC	ACC	GGG	TGA	ACP	LAAC	ACT	'CG1	TGI		. 114	CTCT	
661	S	G	P	С	Ε	G	D	Y	G	G	P	L	V	С	Ε	Q	H	K	M	R	680
2041																	+			TATT + CATAA	2100
681			L		V	I	V	P	G	R	G	С	A	Ι	P	N	R	P	G	I	700
2101																				AGGTA + ICCAT	2160
701	F	V	R	V	A		Y	A	K	W	I	Н	K	I	I	L	T	Y	K		720
2161			AGTO	=	==	217	72														
	_	_	_	4.		70	,														

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#### FIG 1b

1																				GCGC	60
1	TA	CCC	CAC	CGA	GGG	TGA	GGÀ	CGA	CGA	AGA	CTG.	AGT	TAC	GAA	TCC	CCA	GGG	ACC	CGT	CGCG	
1	M	G	W	L	ņ	L	L	L	L	L	Т	Q	С	L	G	V	P	G	Q	R.	20
61																				IGCG ACGC	120
21	S	p P	l AA L	N	D D	GAA F	Q	V			G	T	E	L	Q	Н	L	L	н	A	40
121											<b></b>									CTGT + GACA	180
41		v		G	P	W	Q	E	D	V		D	A	Ξ	Ξ	С	A	G	R	С	60
181							+							. — —						ACTG TGAC	240
61	G	Ď	L	М	D	С	R	A	Ξ	H	Y	И	V	S	S	Н	G	С	Q	L	80
241																				CCTC GGAG	300
81	L	P	W	Т	Q	Н	S	P	H	T	R	L	R	R	S	G	R	С	D	L	100
301							:													GGGC + CCCCG	360
101	F	Q	K	K	D	Y	V	R	T	С	I	M	И	И	G	V	G	Y	3.	G	120
361								L			-+			+-						GAAT GCTTA	420
121	T	М	A	Т	Т	V	G	G	L	P	С	Q	A	W	S	Н	K	Ē	5.	8	140
421								<u> </u>						+				+		ACCCT IGGGA	480
141	D	Ε	K	Y	T	5	Ţ	L	R	N	G	L	Ε	Ε	И	Ē	С	R	N	J,	160
																		(co	ntin	ued)	

(continued)

W	O 99/3	3896	7															PC	T/EF	99/004	78
	(co	ntir	ıue	d)							-	6/	19								
481				+			÷				+			_+-						GAGC  CTCG	540
161			D	P	G	G	p	W	С	Y	T	T	D	P	A	V	R	F	Q	S	180
531				÷			+				+			-+-			+			CCGC  GGCG	600
181			I	K	s	С	R	Ε	А	A	С	v	W	С	N	G	Ε	Ε	Y	R	200
601				<del>+-</del> -			+				+			-+-			+			CCCG  GGGC	660
201	G 1	Ą.	V	D	R	T	Ε	S	G	R	Ε	С	Q	R	W	D	L	Q	H	'n	220
661				+	~		+				+			-+-			+			TTGC  AACG	720
221	н (	2	H	P	F	Ε	P	G	K	ני	L	D	Q	G	L	D	D	И	ž	С	240
721				+			+				+			-+-			+			GCGA + CGCT	780
241	R 1	N .	2	D	G	S	E	R	pı	W	С	ĭ	T	T	D	P	Q	I	Ε	R	260
781				+			+				<del>+</del>			_÷-			+			AACT + TTGA	840
261	E I	F (	С	D	Ŀ	P	R	С	G	S	Ε	A	Q	ņ	R	Q	Ε	Ą	Ţ	T	280
841				<del></del> -			+				<del></del>			-+-			+			TGCG ACGC	900
281	V S	S (	С	F	R	G	K	G	Ε	G	Y	R	G	T	A	N	Ţ	T	Ç	A	300
901				+			+				+			-+-			+			AGAA TCTT	960
301	G 1	<b>V</b>	₽	С	Q	R	W	D	A	Q	I	Þ	Н	Q	Н	R	F	T	D,	Ε	320
961				+			+				+			-÷-			+			GGCG CCGC	1020
321	K Y	Y I	A.	С	К	D	L	R	Ε	N	ш	С	R	И	P.	D	G	S	Ξ	A	340
1021				<b></b>							+									GCGT	1080
341	b A	W (	С	Ξ	T	Ŀ	2,	'n	G	M	R	Ą	A	Ē	С	ž	Q	Ι	D,	R	360
1081				<del></del> -							÷						+			:CGGC :GCCG	1140
361	C :	r	D	D	V	R	P	Q	D	С	ž	H	G	A	G	Ε	Q	¥	R	G	380

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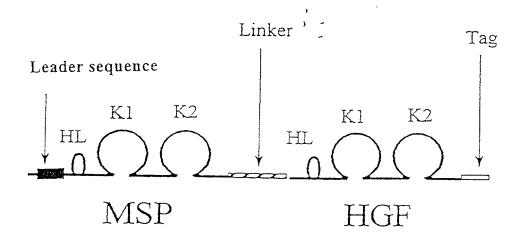
1141	ACGGTCAGCAAGACCCGCAAGGGTGTCCAGTGCCAGCGCTGGTCCGCTGAGACGCCGCAC TGCCAGTCGTTCTGGGCGTTCCCACAGGTCACGGTCGCGACCAGGCGACTCTGCGGCGTG	1200
381	T V S K T R K G V Q C Q R W S A E T P H	400
1201	AAGCCGCAGTTCACGTTTACCTCCGAACCGCATGCACAACTGGAGGAGAACTTCTGCCGG	1260
401	K P Q F T F T S E P H A Q L E E N F C R	420
1261	AACCCAGATGGGGATAGCCATGGGCCCTGGTGCTACACGATGGACCCAAAGGACCCCATTC TTGGGTCTACCCCTATCGGTACCCGGGACCACGATGTGCTACCTGGGTTCCTGGGGTAAG	1320
421	N P D G D S H G P W C Y T M D P R T P F	440
1321	GACTACTGTGCCCTGCGACGCTGCTGATGACCAGCCGCCATCAATCCTGGACCCCCCA 	1380
441	DYCALRRCADDQPPSILDPP	460
1381	GACCAGGTGCAGTTTGAGAAGTGTGGCAAGAGGGTGGATCGGCTGGATCAGCGGCGTTCC+ CTGGTCCACGTCAAACTCTTCACACCGTTCTCCCACCTAGCCGACCTAGTCGCCGCAAGG	1440
461	DQVQFEKCGKRVDRLDQRRS	480
1441	AAGCTGCGCGTGGTTGGGGGCCATCCGGGCAACTCACCCTGGACAGTCAGCTTGCGGAAT TTCGACGCGCACCAACCCCCGGTAGGCCCGTTGAGTGGGACCTGTCAGTCGAACGCCTTA	1500
481	K L R V V G G H P G N S P W T V S L R N	500
1501 501	CGGCAGGCCAGCATTTCTGCGGGGGGTCTCTAGTGAAGGAGCAGTGGATACTGACTG	1560 520
552		320
1561	CGGCAGTGCTTCTCCTCCTGCCATATGCCTCTCACGGGCTATGAGGTATGGTTGGGCACC+++++	1620
521	RQCFSSCHMPLTGYEVWLGT	540
1621	CTGTTCCAGAACCCACAGCATGGAGAGCCAAGCCTACAGCGGGTCCCAGTAGCCAAGATG	1680
541	L F Q N P Q H G E P S L Q R V P V A K M	560
1681	GTGTGTGGGCCCTCAGGCTCCCAGCTTGTCCTGCTCAAGCTGGAGAGATCTGTGACCCTG CACACACCCGGGAGTCCGAGGGTCGAACAGGACGAGTTCGACCTCTCTAGACACTGGGAC	1740
561	V C G P S G S Q L V L L K L E R S V T L	580
1741	AACCAGCGTGTGGCCCTGATCTGCCCCCCTGAATGGTATGTGGTGCCTCCAGGGACC TTGGTCGCACACCGGGACTAGACGGACGGGGGACTTACCATACACCACGGAGGTCCCTGG	1800

\*

# (continued)

	AA	GTG	TGA	GAT	TGC	AGG	CTG	GGG	TGA	GAC	CAA	AGG	TAC	GGG	TAA	TGA	CAC	AGT	CCT.	TAAA	1860
1801	TI	CAC	ACT	CTA	ACG	TCC	GAC	CCC	ACT	CTG	GTT	TCC	ATG	CCC	ATT	ACT	GTG	TCA	GGA	TTTA	1000
601	K	С	Ε	I	A	G	W	G	Ξ	T	K	G	T	G	И	D	T	V	L	И	620
1861				-+-			+			. – – –	+			-+-			+			TGTG + ACAC	1920
621	V	A	£	L	N	V	I	s	N	Q	Ξ	С	N	I	K	H	R	G	R	V	640
1921				-+-			+				+						+			TGAC ACTG	1980
641	R	Ξ	S	Ε	М	С	Ţ	Ε	G	L	Ĭ	A	٥,	V	G	A	С	Ξ	G	D	660
1981				-+-			+				<u> </u>			-+-			+			AATC TTAG	2040
661	Ϋ́	G	G	P	L	A	С	F	T	H	И	С	W	V	L	E	G	I	I	I	680
2041				-+-			+				+			+-			+			GTTT + CAAA	2100
681	P	И	R	V	С	A	R	S	R	M	Ρ	A	V	F	T	R	V	S	V	F	700
2101										GACT CTGA	<del></del>	=		213	16						
701	* *	_	T.T	~	7.7	7,0	T. 7	3.7		τ		+		777							

FIG 2a



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# FIG 2b

1	GAA						1			+ -			TCT				-+	4GG(			60
<u>.</u>	CTT	AAG	GTG	GTA	CCC	CAC	CGA	GGG'	TGA	GGA(	CGA	CGA	AGA(	CTG	AGT:	raco	GAA?	rcc	CAC	3GG	
1				M	G	W	L	2	L	L	L	L	L	Т	Q	С	L	G	V	P	17
61										+			CCG	Ļ ·			-+				120
01	ACC	CGT	CGC	ĠAG	CGG	TAA	CTT.	ACT	GAA	GGT'	TCA	CGA	.GGC	CCC	GTG'	TCT	CGA'	rgr(	CGT	GGA.	
13	G	Q	R	S	D,	L	И	D	F	Q	V	L	'R ]	G	T	Ε	L	Q	H	L	37
101	GCT	ACA	TGC	ĢGT	GGT	GCC	CGG	GCC	TTG	GCA	GGA	GGA	TGT	GGC.	AGA	TGC	TGA.	AGA(	GTG	rgc	180
121	CGA	TGT	ACG	CCA	.CCA	.CGG	GCC	CGG	AAC	CGT	CCT	CCT	'ACA	CCG	TCT.	ACG.	ACT	TCT	CAC	ACG	
38	L	H	A	V	V	P	G	ā	W	Q	E	D	V	A	D	A	Ε	Ε	С	A	57
	TGG	TCG	CTG	TGG	GCC	CTT	'AAT	GGA	.CTG	CCĢ	GGC	CTI	CCA	CTA	CAA	CGT	GAG	CAG	CCA	TGG	240
181	ACC	AGC	GAC	ACC	CGG	GAA	TTA	CCT	GAC	GGC	CCG	GAA	GGT	GAT	GTT	GCA	.CTC	GTC	GGT.	ACC	
58	G	R	С	G	p	L	М	D	С	R	А	F	H	Y	N	V	S	S	H	G	77
0.43							,						ACAC	+							300
241	AAC	GGT	TGA	CG2	CGG	TAC	CTG	AGT	TGI	'GAG	CGG	GG3	GTG	CTC	:CGA	CGC	CGC	AAG	ACC	CGC	
78	С	Q	Ľ	L	P	W	T	Q	Н	S	p	H	T	R	Ţ	R	R	S	G	R	97
201	CTC	STGA	CCI	CTI	CCF	AGA.P	AGAA	AGA	CTA	CGI	ACG	GAC	CTG	CAT	CAT	'GAA	CAA	TGG	GGT	TGG	360
301	GAC	ACI	:GG?	,GA,	\GG"	CTI	CTI	TCI	GAI	GC.	TGC	CTC	<b>GAC</b>	GTA	AGTA	CTI	GTT	'ACC	CCA	ACC	
98	С	G	L	F	Q	K	K	D	ž	V	R	T	С	I	М	N	N	G	V	G	117
٠													rgco				+-				420
361	CAT	rggc		GTO	GGT?	ACCO	GTC	CTC	GCF	ACC(	CACC	CGG	ACGO	ĠAC	CGG1	CCC	SAAC	CTC	GGT	GTT	
118	Y	R	G	T	M	А	T	Т	V	G	G	L	₽	С	Q	A	W	S	H	K	137
	GT:	rcco	CGAZ	ATG!	ATC	ACA	AGŢ <i>I</i>	ACA(	CGC	CAC	CTC	rcc	GGAZ	ATGO	GCC:	rgg:	AAG <i>I</i>	AGA	CTI	CTG	480
421	CA	AGG(	GCT	-+-: rac:	rag'	TGT	rca:	GTO	GCG(	GGT(	JAG?	₹GG	CCT	CAC	CGG	ACC:	rtĊī	CTI	[GA#	AGAC	
138	F	P	N	D	Н	К	Y	Ť	7.	Ţ	L	R	И	G	Ľ	Ε	E	N	Ē	С	157
	CC	GTA	ACC	CTG	ATG	GCG2	ACC	CCG	GAG(	GTC	CTT	GGT	GCT	ACA(	CAA	CAG.	ACC	CTG	CTG:	rgcg	540
481																				ACGC	240
158													Y								177

(continued)

(continued)

WO 99/38967 PCT/EP99/00478 (continued) 11/19 CTTCCAGAGCTGCGGCATCAAATCCTGCCGGGAGGCCGCGTGTGTCTGGTGCAATGGCGA 600 541 GAAGGTCTCGACGCCGTAGTTTAGGACGGCCCTCCGGCGCACACAGACCACGTTACCGCT F Q S C G I K S C R E A A C V W C 178 197 GGAATACCGCGGCGCGGTAGACCGCACGGAGTCAGGGCGCGAGTGCCAGCGCTGGGATCT 601 CCTTATGGCGCCGCCCATCTGGCGTGCCTCAGTCCCGCGCTCACGGTCGCGACCCTAGA EYRGAVDRTESGRECQRWDL 198 217 TCAGCACCCGCACCAGCACCCCTTCGAGCCGGGCAAGTTCCTCGACCAAGGTCTGGACGA 661 720 AGTCGTGGGCGTGGTCGTGGGGAAGCTCGGCCCGTTCAAGGAGCTGGTTCCAGACCTGCT Q H P H Q H P F E P G K F L D Q G L D D 218 237 CAACTATTGCCGGAATCCTGACGGCTCCGAGCGGCCATGGTGCTACACTACGGATCCGCA 721 780 GTTGATAACGGCCTTAGGACTGCCGAGGCTCGCCGGTACCACGATGTGATGCCTAGGCGT NYCRNPDGSERPWCYTTDPO 238 257 GATCGAGCGAGAGTTCTGTGACCTCCCCCGCTGCGGGTCCGAGGCACAGCCCCGCCTCGA 781 840 I E R E F C D L P R C G S E A Q P R L 258 277 GGGCGGTGGCGGTTCTCGGTGGCGGTGGCCGGTGGCGGTTCTCTAGAGGGACAAAG 900 CCCGCCACCGCCAAGACCACCGCCACCGAGGCCGCCAAGAGATCTCCCTGTTTC 297 278 GAAAAGAAGAATACAATTCATGAATTCAAAAAATCAGCAAAGACTACCCTAATCAAAAT 960 901 CTTTTCTTCTTATGTTAAGTACTTAAGTTTTTTAGTCGTTTCTGATGGGATTAGTTTTA I H E FKKSAKT 298 317 AGATCCAGCACTGAAGATAAAAACCAAAAAAGTGAATACTGCAGACCAATGTGCTAATAG 1020 961 TCTAGGTCGTGACTTCTATTTTTGGTTTTTTCACTTATGACGTCTGGTTACACGATTATC D P A L K I K T K K V N T 337 318 ADQCANR ATGTACTAGGAATAAAGGACTTCCATTCACTTGCAAGGCTTTTGTTTTTGATAAAGCAAG 1080 1021 TACATGATCCTTATTTCCTGAAGGTAAGTGAACGTTCCGAAAACAAAAACTATTTCGTTC CTRNKGLPFTCKAFVFDKAR 338 AAAACAATGCCTCTGGTTCCCCTTCAATAGCATGTCAAGTGGAGTGAAAAAAAGAATTTGG 1140 1081 K Q C L W F P F N S M S S G V K K E F G 358 CCATGAATTTGACCTCTATGAAAACAAGACTACATTAGAAACTGCATCATTGGTAAAGG 1200 1141 GGTACTTAAACTGGAGATACTTTTGTTTCTGATGTAATCTTTGACGTAGTAACCATTTCC HEFDLYENKDYIRNC 397 378

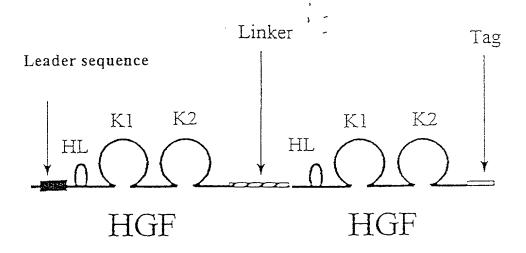
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ACG	CAG	CTA												CAA	ATG	TCA	GCC	CTG	GAG
TGC	GTC	GAT	GTT	CCC.	TTG	TCA	TAG	ATA	GTG	ATT	CTC	ACC	GTA	GTT.	TAC	AGT	'CGG	GAC	CTC
R	S	Y	K	G	T	V	S	I	Ţ	K	S	G	I	K	С	Q	P	W	S
TTC	CAT	GAT.	ACC.	ACA(	CGA	ACA	CAG	CTA	TCG	GGG	TAA	AGA	CCT +	ACA	.GGP	AAA -+-	CTA	.CTG	TCC
AAG	GTA	CTA	TGG'	TGT	GCT	TGT	GTC	GAT	AGC	CCC	TŢĄ	TCT	GGA	TGT	CCI	TTT	GAT	GAC	AG
S	М	I	D,	H	E	Н	S	Y	R	G	K	D	L	Q	Ξ	N	Y	С	R
	TCC		<b>+</b>			-+-			+				+			+-			
N	'AGG		G				G						s s	N.	AG0	,1C1 E		.1 GC R	GA. Y
	AGT TCA		+						+				<b>+</b>			-+-			
Ε	V	С	D	Ι	P	Q	С	S	Ε	V	Ε	С	М	T	С	N	G	Ε	S
	TCG		÷			-+-			+				+			+-			
Y	R	G	L	М	D	Н	T	Ε	S	G	K	I	С	Q	R	W	D	Н	Q
	ACC		÷						+				+			+-			
T	P	Н	R	H	K	ŗ	L	P.	Ε	Ŗ	Ϋ́	P	D	K	G	F	D	D	N
	TTG AAC		+			-+-							· <del></del>			+-			
Y	С	R	N	P	D	G	Q	Ď	R	Ď	W	С	Y	T	L	D	P	Н	T
	CTG GAC		÷			-+-			~							+-			
R	W	Ξ	Ϋ́	С	A	I	K	T	С	A	D	K	A	D	D	D	D	K	Н
	CCA GGT		+			-==	=			17	09								
н			H		H	*			• -	5 6	3								

FIG 3a



. . . . .

# FIG 3b

						+-			_==	F			-+-			+			GCTG + CGAC
	IMG	GCG	المدي	GGG	CAG	310	0:0	3100	M	W	V	T	K	L	L	P	A	L L	L
CT	GCA	GCA	TGT	CCT	CCT	GCA'	TCT	CCT	CCT	GCT		CAT	CGC	CAT	CCC	CTA	TGC	AGA	GGGA
GΑ	CGT	CGT	AĊA	GGA	GGA	CGT	AGA(	GGA(	GGA(	CGA	GGG:		_	GTA	GGG -	GAT	ACG	TCT	CCCT
L	Q	Н	V	L	L.	Н	L	L	·L·	Ľ	P.	Ī	Ā	I	P	Ÿ	A.	E	Ğ.¯
						+				÷			-+-						AATC
GT	TTC	CTT	TTC	TTC	TTT.	ATG'	TTA	AGT.	ACT	TAA	GTT	TTT	TAG	TCG	TTT	CTG	ATG	GGA	TTAG
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	TGTCGAAATCCTCGAGGGGAAGAAGGGGGACCCTGGTGTTTCACAAGCAATCCAGAGGTA	
541	ACAGCTTTAGGAGCTCCCCTTCTTCCCCCTGGGACCACAAGTGTTCGTTAGGTCTCCAT	600
172	C R N P R G E E G G P W C F T S N P E V	191
601	CGCTACGAAGTCTGTGACATTCCTCAGTGTTCAGAAGTTGAATGCATGACCTGCAATGGG GCGATGCTTCAGACACTGTAAGGAGTCACAAGTCTTCAACTTACGTACTGGACGTTACCC	660
192	RYEVCDIPQCSEVECMTCNG	211
661	GAGAGTTATCGAGGTCTCATGGATCATACAGAATCAGGGAAGATTTGTCAGCGCTGGGAT CTCTCAATAGCTCCAGAGTACCTAGTATGTCTTAGTCCGTTCTAAACAGTCGCGACCCTA	720
212	E S Y R G L M D H T E S G K I C Q R W D	231
721	CATCAGACACCACCGGCACAATTCTTGCCTGAAAGATATCCCGACAAGGGCTTTGAT GTAGTCTGTGGTGGCCGTGTTTAAGAACGGACTTTCTATAGGGCTGTTCCCGAAACTA	780
232	H Q T P H R H K F L P E R Y P D K G F D	251
781	GATAATTATTGCCGCAATCCCGATGGCCAGCCGAGGCCATGGTGCTATACTCTTGACCCT CTATTAATAACGGCGTTAGGGCTACCGGTCGGCTCCGGTACCACGATATGAGAACTGGGA	840
252	D N Y C R N P D G Q P R P W C Y T L D P	271
841	CACACCCGCTGGGAGTACTGTGCAATTAAAACATGCGCTGACAAAGCTTCGGGCGGTGGC GTGTGGGCGACCCTCATGACACGTTAATTTTGTACGCGACTGTTTCGAAGCCCGCCACCG	900
272	H T R W E Y C A I K T C A D K A S G G G	291
901	GGTTCTGGTGGCGGTGGCTCCGGCGGTGGCGGTTCTCTAGAGGGACAAAGGAAAAGAAGA CCAAGACCACCGCCACCGAGGCCGCCACCGCCAAGAGATCTCCCTGTTTCCTTTCT	960
292	G S G G G G G G S L E G Q R K R R	311
961	AATACAATTCATGAATTCAAAAAATCAGCAAAGACTACCCTAATCAAAATAGATCCAGCA TTATGTTAAGTACTTAAGTTTTTTAGTCGTTTCTGATGGGATTAGTTTTATCTAGGTCGT	1020
312	NTIHEFKKSAKTTLIKIDPA	331
1021	CTGAAGATAAAAACCAAAAAAGTGAATACTGCAGACCAATGTGCTAATAGATGTACTAGG GACTTCTATTTTTGGTTTTTTCACTTATGACGTCTGGTTACACGATTATCTACATGATCC	1080
332	LKIKTKKVNTADQCANRCTR	351
1081	AATAAAGGACTTCCATTCACTTGCAAGGCTTTTGTTTTTGATAAAGCAAGAAAACAATGC TTATTTCCTGAAGGTAAGTGAACGTTCCGAAAACAAAAACTATTTCSTTCTTTTGTTACG	1140
352	N K G L P F T C K A F V F D K A R K Q C	371
1141	CTCTGGTTCCCCTTCAATAGCATGTCAAGTGGAGTGAAAAAAGAATTTGGCCATGAATTT GAGACCAAGGGGAAGTTATCGTACAGTTCACCTCACTTTTTTCTTAAACCGGTACTTAAA	1200
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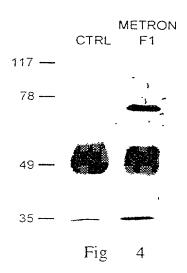
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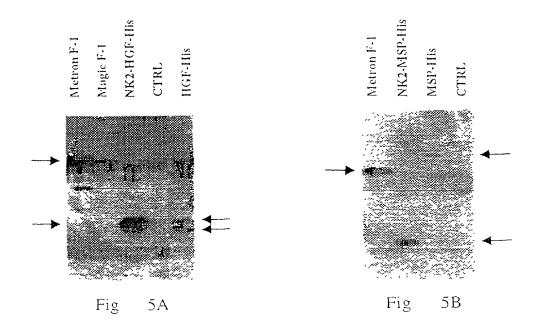
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1261	ΑA	.GGC	AAC	AGI	ATC	TAI	CAC	TAP	\GAG	TGG	CAT	'CAA	ATC	TCA	.GCC	CTG	GAG	TTC	CAT	GATA	
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1321	CC	ACA	LCGA	ACA	CAC	CTA	TCG	GGG	TAA	LAGA	ÇCT	'ACA	.GGA	- AAA	.CTA	.CTG	TCG	AAA	TCC	TCGA	
	GG	TGI	GCI	TGT	GTC	GAT	'AGC	CCC	ATT	TCT	'GGA	TGT	CCI	TTT	GAI	GAC	AGC	TTT	AGG	AGCT	13
432	P	H	Ε	H	S	Y	R	G	K	D	L	Q	Ε	N	Y	С	R	N	P	R	45
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1301	CC	CCI	TCT	TCC	CCC	TGG	GAC	CAC	AAA	GTG	TTC	GTT	AGG	TCT	CCA	TGC	GAT	GCT	TCA	GACA	1.4
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1621	AA	TCC	CGA	TGG	CCA	GCC	GAG	GCC	ATG	GTG	CTA	TAC	TCT	TGA	ccc	TCA	CAC	CCG	CTG	GGAG	
	TT.	AGG	GCT.	ACC	GGT	CGG	CTC	CGG	TAC	CAC	GAT	ATG	AGA	ACT	GGG	agt Agt	GTG	GGC	GAC	CCTC	16
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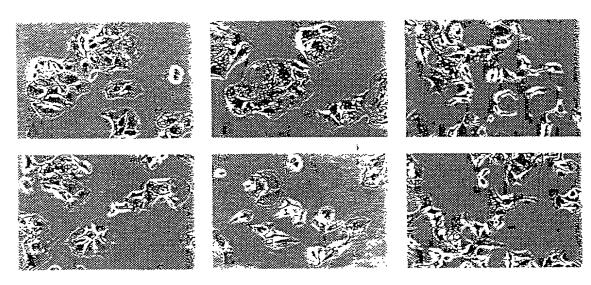
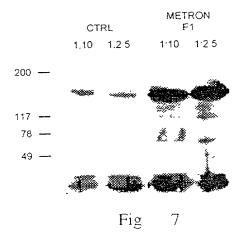
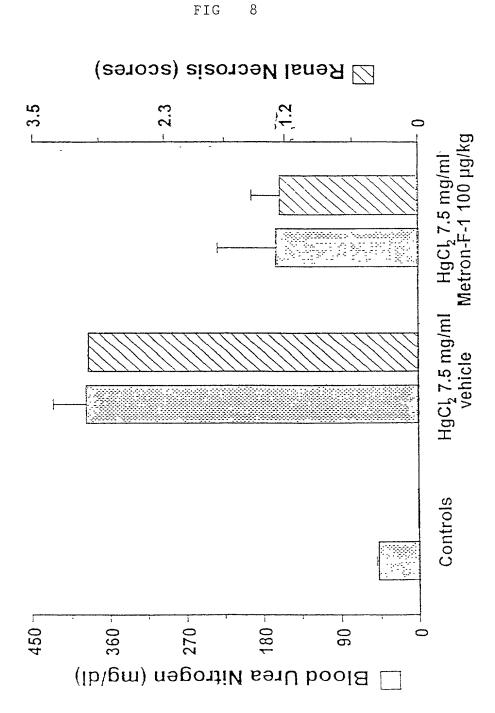


Fig 6





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Attorney Docket No.:

471-162P

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# COMBINED DECLARATION AND POWER OF ATTORNEY FOR PATENT AND DESIGN APPLICATIONS

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated next to my name; that I verily believe that I am the original, first and sole inventor (if only one inventor is named below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Insert Title:	<u>Recombinant p</u>	<u>roteins</u>	derived from	HGF and MSP			
Fill in Appropriate	the specification of w	hich is attache	d hereto. If not attacl	ned hereto,			as
Information - For Use Without	the specification	was filed on _	oer	00478			;
Specification	and amended on	meddon mani			(if appli	icable) a	and/or
Attached:	the specification	was filed on _	27.01.1999	00770			as PCT
	International App	lication Numb	er <u>PCT/EP99/</u>	004/8		if appl	icable)
	amended under F	CI Article 19	on	00478		ind on	ocification
The state of the s	including the claims, I acknowledge th of Federal Regulation I do not know an	as amended by e duty to disc s, §1.56. d do not belie	y any amendment ref lose information which we the same was ever	known or used in the Uny printed publication olication, that the same this application, that the same this application of this application, and that no application, and that no application foreign to the Unite except as follows.	bility as defined nited States of A	l in Tit	le 37, Code before my
Section 19	the United States of A	merica on an	application filed by r	ne or my legal represent	ative or assigns	more 1	than twelve
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	I hereby claim	foreign priori	ty benefits under T	itle 35, United States	Code, §119(a)-( so identified b	a) or a elow a	iny foreign
mora files end	application(s) for pa	t or inventor's	certificate having a	filing date before that of	the application	on wh	ich priority
Analysis of the second of the	is claimed:		· ·	-			
100 To 10	Prior Foreign Appli	cation(s)			Pric	rity Cl	laimed
Insert Priority	0			30.01.1998		X	
	MI98A000179	<u>  Ltaly</u> (Country)		(Month/Day/Year Filed	4)	Yes	No
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	(IATHIDCI)	(Country)		, , ,,			
	(Number)	(Country)		(Month/Day/Year File	<u>d)</u>	Yes	No
	I hereby claim the applications(s) listed	benefit und	er Title 35, United	States Code, §119(e) o	of any United	States	provisional
Insert Provisional Application(s): (if any)	(Application Number	)		(Filing Date)			
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	All Foreign Applicati Designs) Prior to the	ons, if any, fo Filing Date of	or any Patent or Inver This Application:	ntor's Certificate Filed M	ore than 12 Mo	nths (6	Months for
	Country		Application Number	Date of F	iling (Month/Da	y/Year)	
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	code, §112, I acknown	nd/or PC1 appowledge the d	uty to disclose informations 61.56 which	Code, §120 of any United of the claims of this er provided by the first provided by the first provided is material became available betward of this application.	ed States and/o application is n paragraph of Tit I to the patenta veen the filing	r PCT a ot disc le 35, l bility a date o	pplication(s) losed in the Jnited States is defined ir of the prior
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	(Application Numbe	r)	(Filing Date)	(Status -	patented, pendi	ng, aba	ndoned)
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#### Attorney Docket No.:

I hereby appoint the following attorneys to prosecute this application and/or an international application based on this application and to transact all business in the Patent and Trademark Office connected therewith and in connection with the resulting patent based on instructions received from the entity who first sent the application papers to the attorneys identified below, unless the inventor(s) or assignee provides said attorneys with a written notice to the contrary:

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the

validity of the application or any patent issu	ied thereon.		
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Enzo MEDICO	1 100		
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Gianfrânco CASELLI	6-6-6-1		27.07.2000
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L'AQUILA, Italy	<del>~</del> <del>/</del>	Italia	n
POST OFFICE ADDRESS (Complete Street Ad	ldress including City, State & Co	ountry)	
Via Campo di Pile - L'AQ	UILA, Italy		
GIVEN NAME/FAMILY NAME	INVENTOR'S SIGNATURE		DATE*
Paolo COMOGLIO	1		27.07.2000
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POST OFFICE ADDRESS (Complete Street Ad		ountry)	
Via Campo di Pile - L'AQ	UILA, Italy		

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Full Name of Second Inventor, if any: See above

Full Name of Third Inventor, if any: see above

Full Name of Fourth Inventor, if any: see above

Full Name of Fifth Inventor, if any: see above